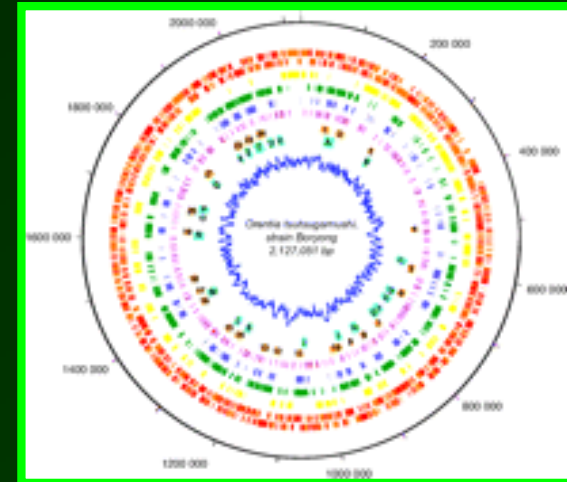
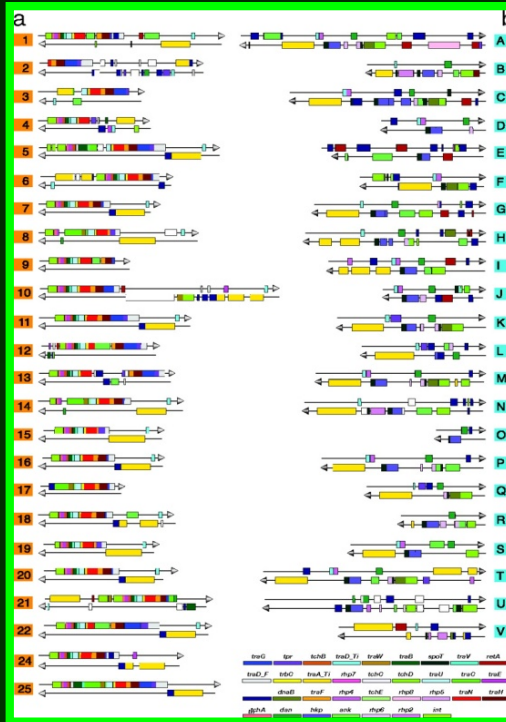
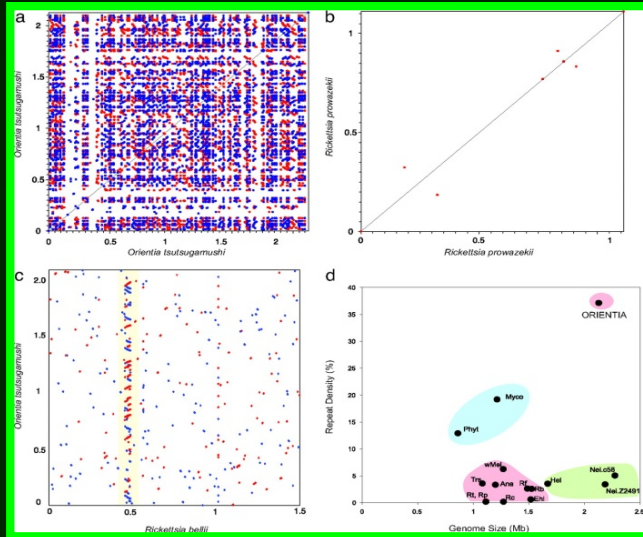


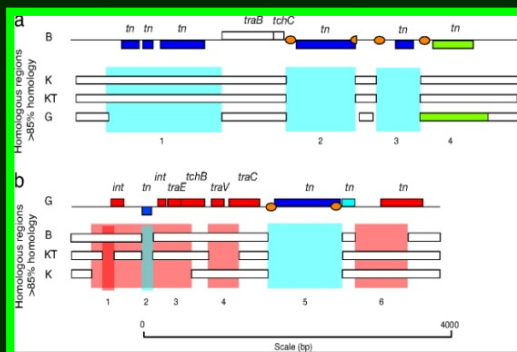
*Diversity of Orientia tsutsugamushi strains
in South East Asia (Thailand, Cambodia,
Vietnam) based on the 56-kDa protein gene
and multilocus typing*

Roger Frutos
Cirad, Montpellier

Orientia tsutsugamushi genome structure



ca. 2 Mb
High repeat number
Low coding percentage

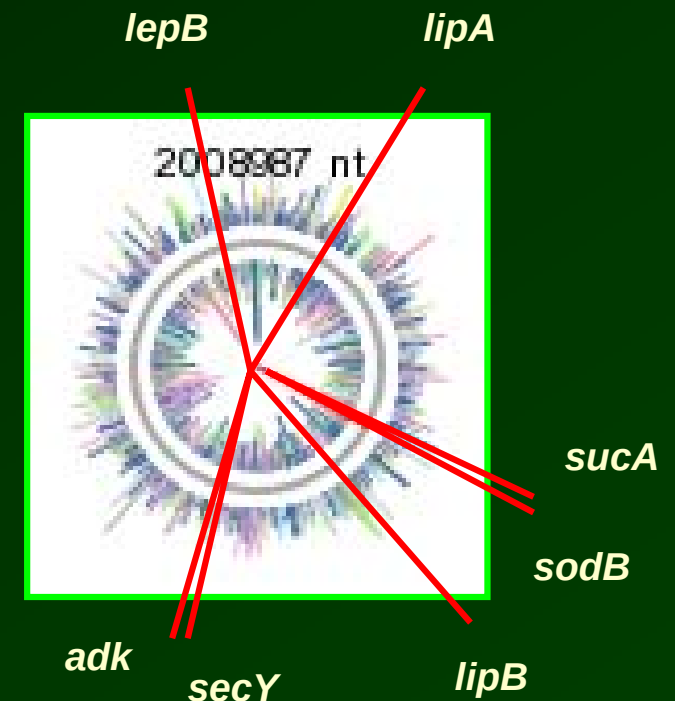


Gene duplication
High pseudogene contents
High number of mobile elements
Highly repeated tra sequences 'type IV secretory pathway'
Limited core genome

MLST target loci

7 target genes

<i>adk</i>	adenosine kinase
<i>lepB</i>	GTP-binding protein
<i>lipA</i>	lipoyl synthase
<i>lipB</i>	lipoyltransferase
<i>secY</i>	preprotein translocase
<i>sodB</i>	Superoxide dismutase
<i>sucA</i>	2-oxoglutarate dehydrogenase



Validated on other Rickettsiales (*Ehrlichia ruminantium*)
Part of the core genome

Strains

Region-wide analysis

MLST : Cambodia (Duong et al., 2010 – this presentation)

Thailand : under way

Vietnam : under way

56-kDa protein gene

Cambodia - Thailand - Vietnam

Combination of several works

Patients diagnosed with scrub typhus

Strains pathogenics for humans

2 reference strains from Korea and Japan

Goal: have an idea of the distribution, diversity and dynamics at the scale of the South East Asia region

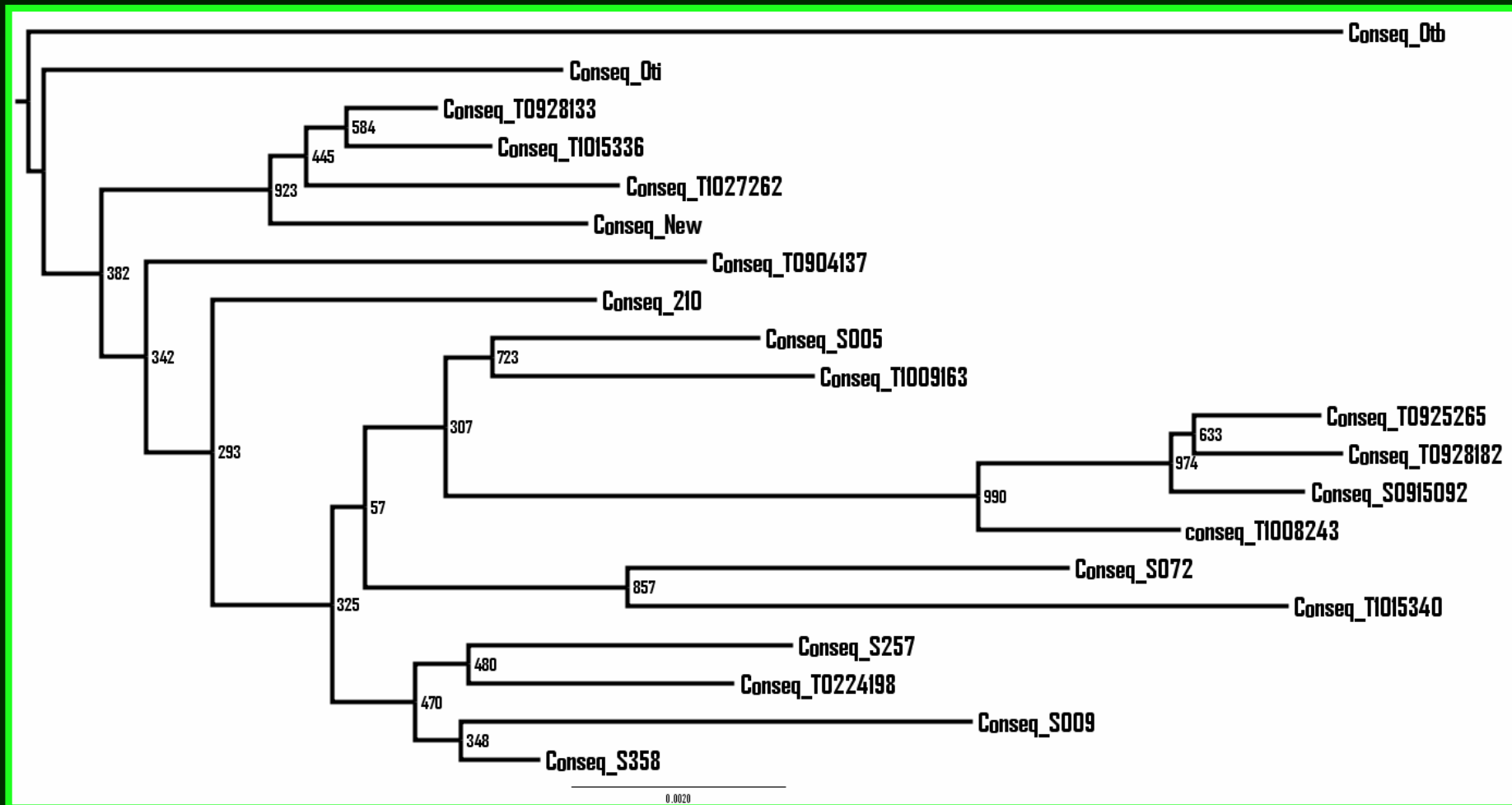
MLST

	<i>adk</i>	<i>lepB</i>	<i>lipA</i>	<i>lipB</i>	<i>secY</i>	<i>sodB</i>	<i>sucA</i>
Strain 005	1	1	1	1	1	1	1
Strain 009	2	2	2	2	2	2	2
Strain 072	3	3	3	3	2	3	3
Strain 210	4	4	4	4	3	4	4
Strain 257	5	5	5	5	2	5	5
Strain 358	6	6	2	6	2	2	4
Strain New	7	7	2	7	3	6	4
Strain Otb	8	8	6	1	4	7	6
Strain Oti	9	9	7	8	5	8	7
Strain S0915092	10	10	8	4	6	9	8
Strain T0224198	6	6	9	9	2	10	4
Strain T0904137	6	11	10	10	7	11	9
Strain T0925265	11	12	11	11	6	12	4
Strain T0928133	12	7	2	7	8	13	4
Strain T0928182	10	12	8	11	9	14	8
Strain T1008243	1	12	8	11	6	13	10
Strain T1009163	1	13	4	12	1	15	4
Strain T1015336	12	7	2	7	3	16	8
Strain T1015340	13	14	12	7	2	14	11
Strain T1027262	12	15	13	7	3	13	4
N Haplotypes	13	15	13	12	9	16	11

One patient – One strain – One ST
Clonal structure

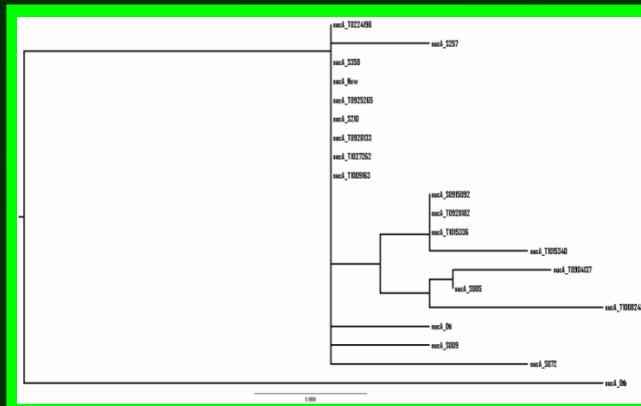
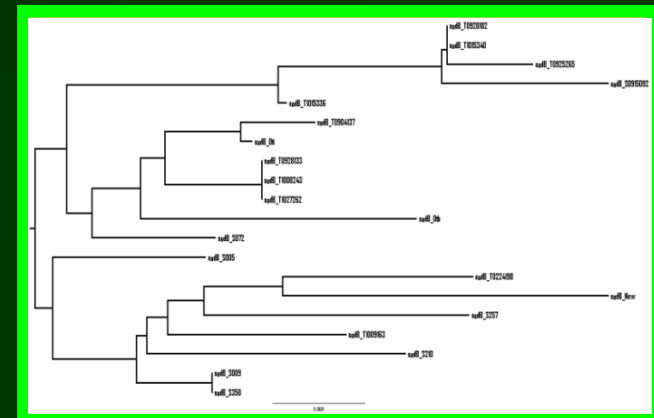
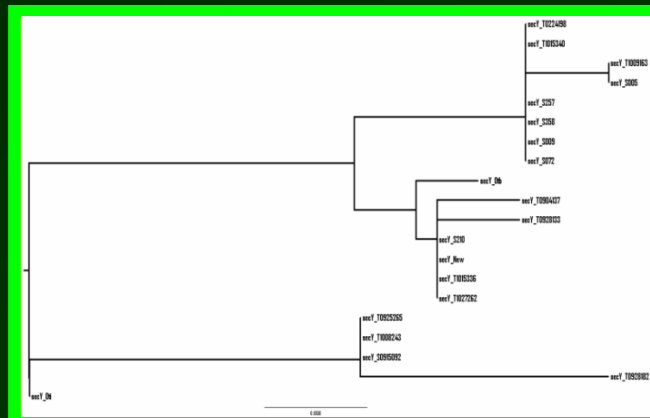
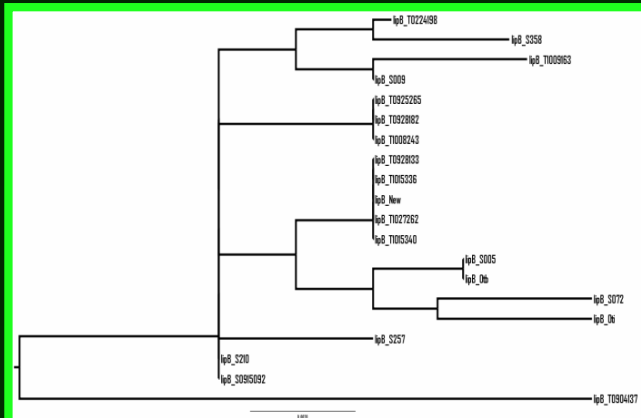
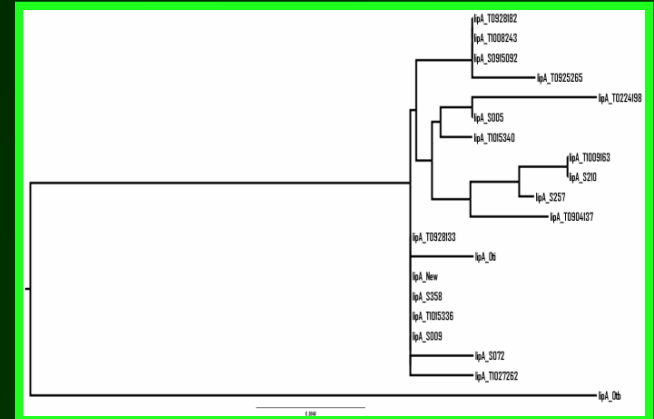
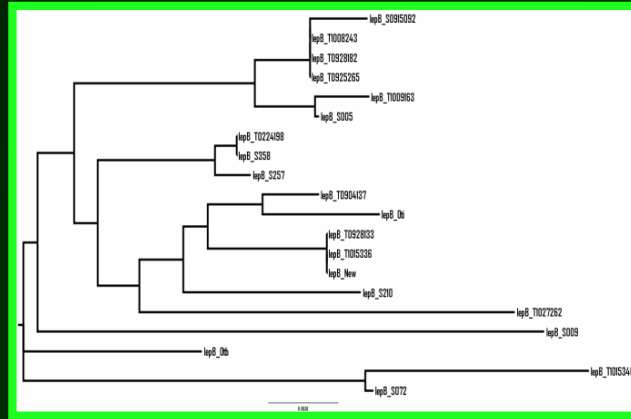
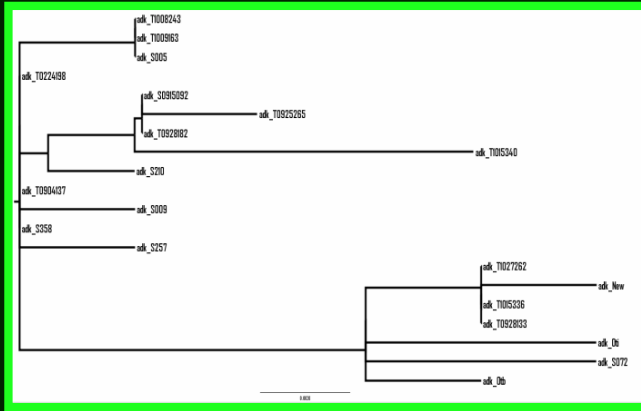
Specific habitat (chigger mites)
Divergent evolutionary paths (literature)
Positive selection

Concatenated sequences



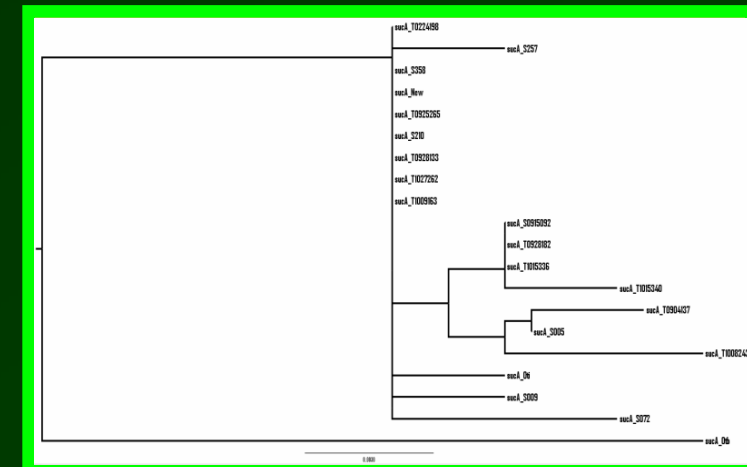
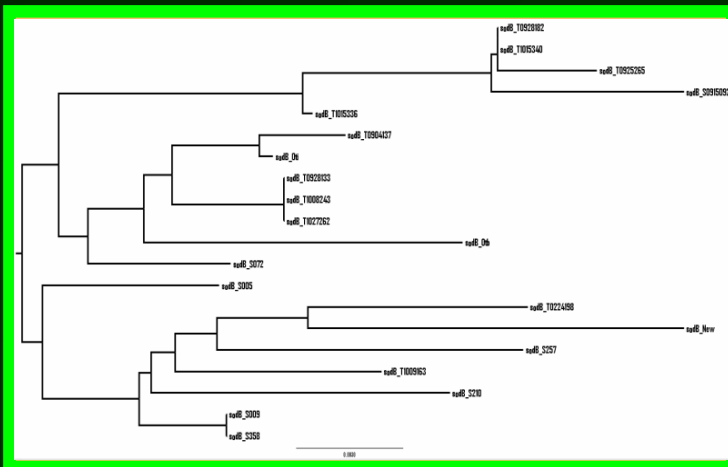
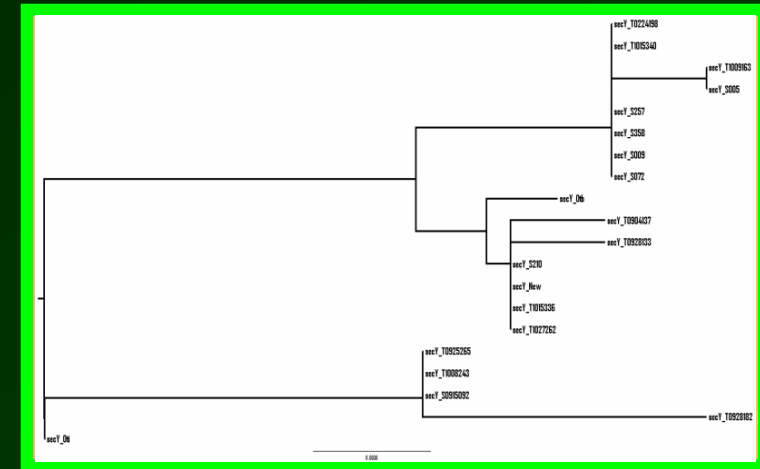
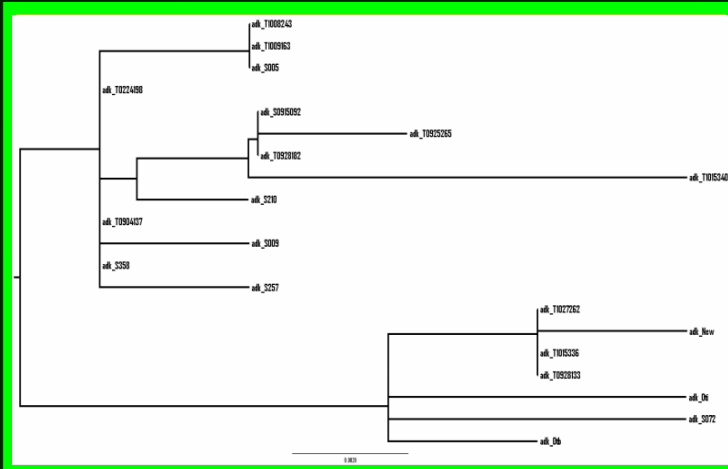
Cambodia vs. Korea/Japan
Weak bootstraps
Not clonal

Individual gene trees



Differing topologies
Different evolutionary paths
Oti and Oti often separate
Not always / not at the same level

Differing topologies



Genes expected to be linked
Differing evolutionary paths

Neutrality

	Hp	S	h	h (s)	Pa	Na	Ns	Tajima's D	D*	F*
<i>adk</i>	13	18	19	12	6	10	9	-1.18764 ^{NS}	-2.05288 ^{NS}	-2.09066 ^{NS}
<i>lepB</i>	15	34	36	12	22	11	25	-0.51375 ^{NS}	-0.36526 ^{NS}	-0.47702 ^{NS}
<i>lipA</i>	13	24	26	21	3	9	17	-2.05011*	-3.02265**	-3.18223**
<i>lipB</i>	12	14	14	9	5	8	6	-1.39525 ^{NS}	-1.75136 ^{NS}	-1.91243 ^{NS}
<i>secY</i>	9	17	18	5	12	3	14	0.07808 ^{NS}	-0.19261 ^{NS}	-0.13160 ^{NS}
<i>sodB</i>	16	27	27	12	15	9	18	-0.55983 ^{NS}	-0.81597 ^{NS}	-0.86129 ^{NS}
<i>sucA</i>	11	19	19	17	2	10	9	-2.11308*	-3.16817 ^{NS}	-3.32210 ^{NS}

Impossible to reject the null hypothesis of neutrality

Negative selection of core function genes (not neutral) : No

Positive selection of strains (not neutral) : No

Blurred figures : Not clonal but kind of “amphimictic”

DNA polymorphism

	Hp	S	η	Pa	$\eta(s)$	θ	π
<i>Adk</i>	13	18	19	6	12	5,35	0.00947
<i>lepB</i>	15	34	36	22	12	10.15	0.02191
<i>lipA</i>	13	24	26	3	21	7.33	0.00806
<i>lipB</i>	12	14	14	5	9	9.95	0.00718
<i>secY</i>	9	17	18	12	5	5.07	0.01257
<i>sodB</i>	16	27	27	15	12	7.52	0.01825
<i>sucA</i>	11	19	19	2	17	5.36	0.00619

Recombination and linkage disequilibrium

	Rm	ZnS	B	Q	PW	F	BP
<i>adk</i>	0	0.0863	0,0000	0,0000	136	6**	3
<i>lepB</i>	8	0.1195	0.1290	0.2188	496	33*/**	1
<i>lipA</i>	1	0.3997	0.4899	0.5000	231	0	0
<i>lipB</i>	1	0.1307	0.2308	0.2857	91	0	0
<i>secY</i>	0	0,2660	0.2667	0.4375	120	24*/**	16
<i>sodB</i>	7	0.0906	0.0385	0.0741	351	16*/**	9
<i>sucA</i>	1	0.2257	0.2222	0.2632	171	0	0

Evidence for recombination

Contradictory trends ?

STs

Clonal populations
Isolated patches
Divergent evolutionary paths

DNA polymorphism

Recombination
Exchange of genetic material
Co-infection (intracellular)

Contradiction does not exist in nature

Only things we do not understand yet

Chigger mite: the true host

Vertical transmission

Symbiont-like association

No host to host movement



Small mammals: the maintenance host

Multiple infections

Recombination process

Genetic diversity

Humans: the accidental host

Rare and unique genetic events

Looks clonal but is not



Different works same results

Mahidol Oxford Tropical Medicine Research Unit

Sonthayanon et al. 2010. High rates of homologous recombination in the mite endosymbiont and opportunistic human pathogen *Orientia tsutsugamushi*. PLoS Negl Trop Dis 4, e752.

Pasteur Institute in Cambodia – Cirad –University Montpellier 2

Duong et al., 2010. Diversity of *Orientia tsutsugamushi* clinical isolates in Cambodia reveals active selection and recombination process. Infect Genet Evol. doi:10.1016/j.meegid.2010.08.015

Same approach

Different strains

Different countries

Different loci

SAME RESULTS

SAME CONCLUSIONS

56-kDa type specific antigen (TSA)

High genomic plasticity

High level of duplication and rearrangements

Numerous repeats transposons and conjugative elements

Horizontal transfer

MLST analysis: extensive recombination

56-kDa protein gene

Unique to *O. tsutsugamushi*

Outer membrane surface

Induces neutralizing antibody responses

Involved in cell invasion (specific binding to fibronectin)

Interacts directly with the mammalian host

Subjected to host-driven adaptive selection

9 significant clusters

Good target for investigating population structure

56-kDa TSA diversity in Thailand

Variable domain I (VD I) to domain IV (VD IV)
PCR amplification and sequencing

Country-wide sampling scheme (63 strains)
8 genotypes reported previously
Only 4 detected here

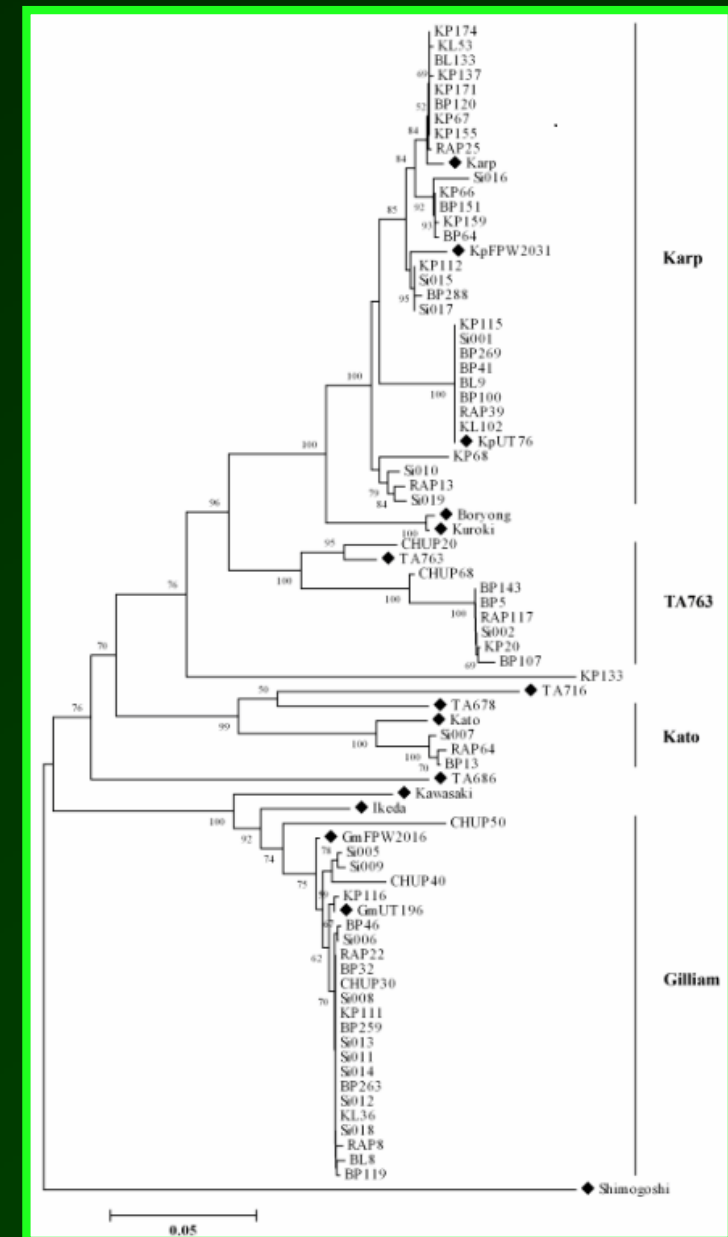
Karp: 46%

Gilliam: 34.9%

TA763: 12.7%

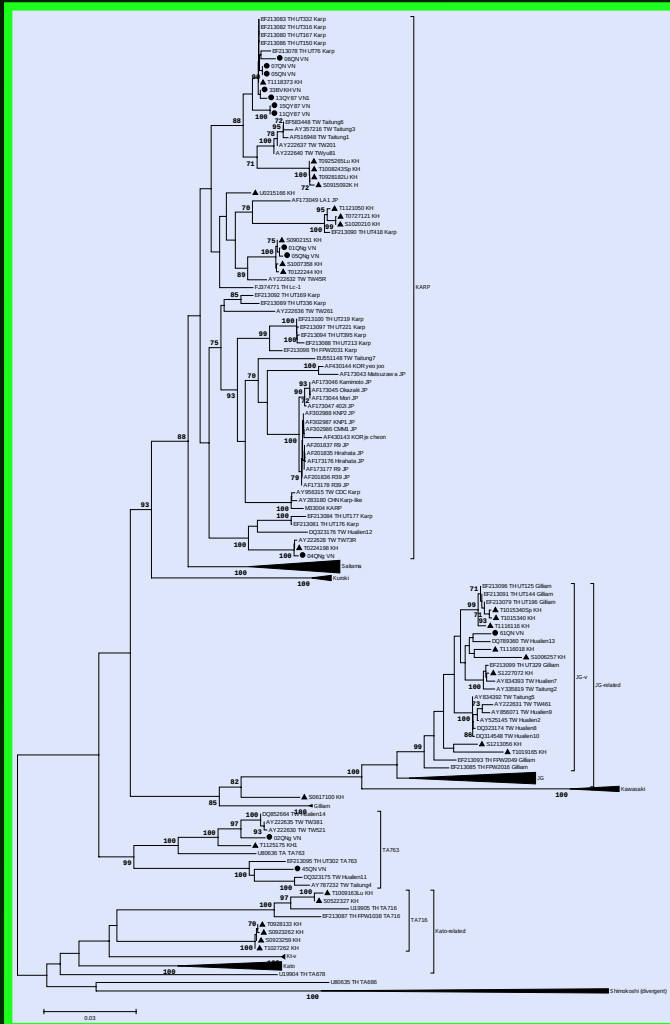
Kato: 4.8%

Karp genotype confirmed as the most common
Gilliam and TA763 also commonly found (contrast)
TA716 was not found
Difference with previous reports
Spatial and temporal variations ?



56-kDa TSA diversity in Cambodia and Vietnam

Full length sequence PCR amplification and sequencing



Vietnam

3 groups

Karp (77%)
TA763 (15.5%)
JG-v (7.5%)

Cambodia

5 major groups

**Karp (43.5%),
JGv (25%)
Kato/TA716 (21.5%)
TA763 (3.5%)
Gilliam (3.5%)**

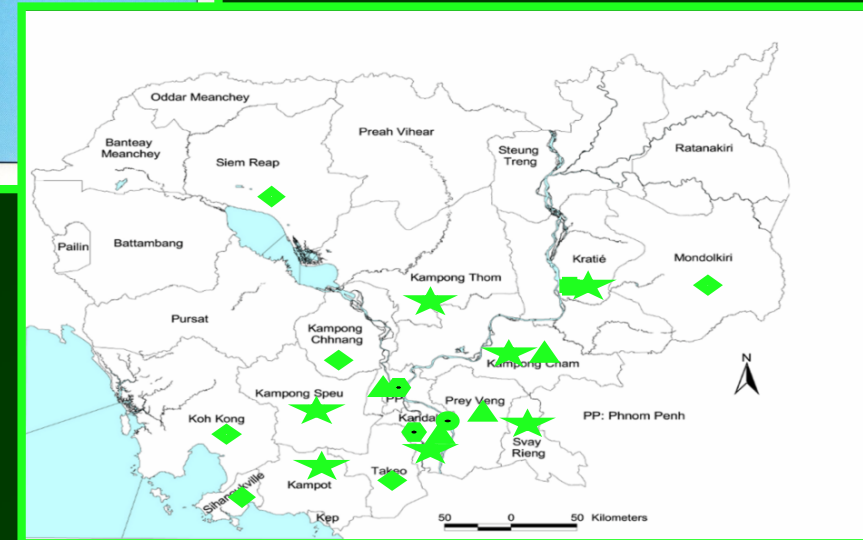


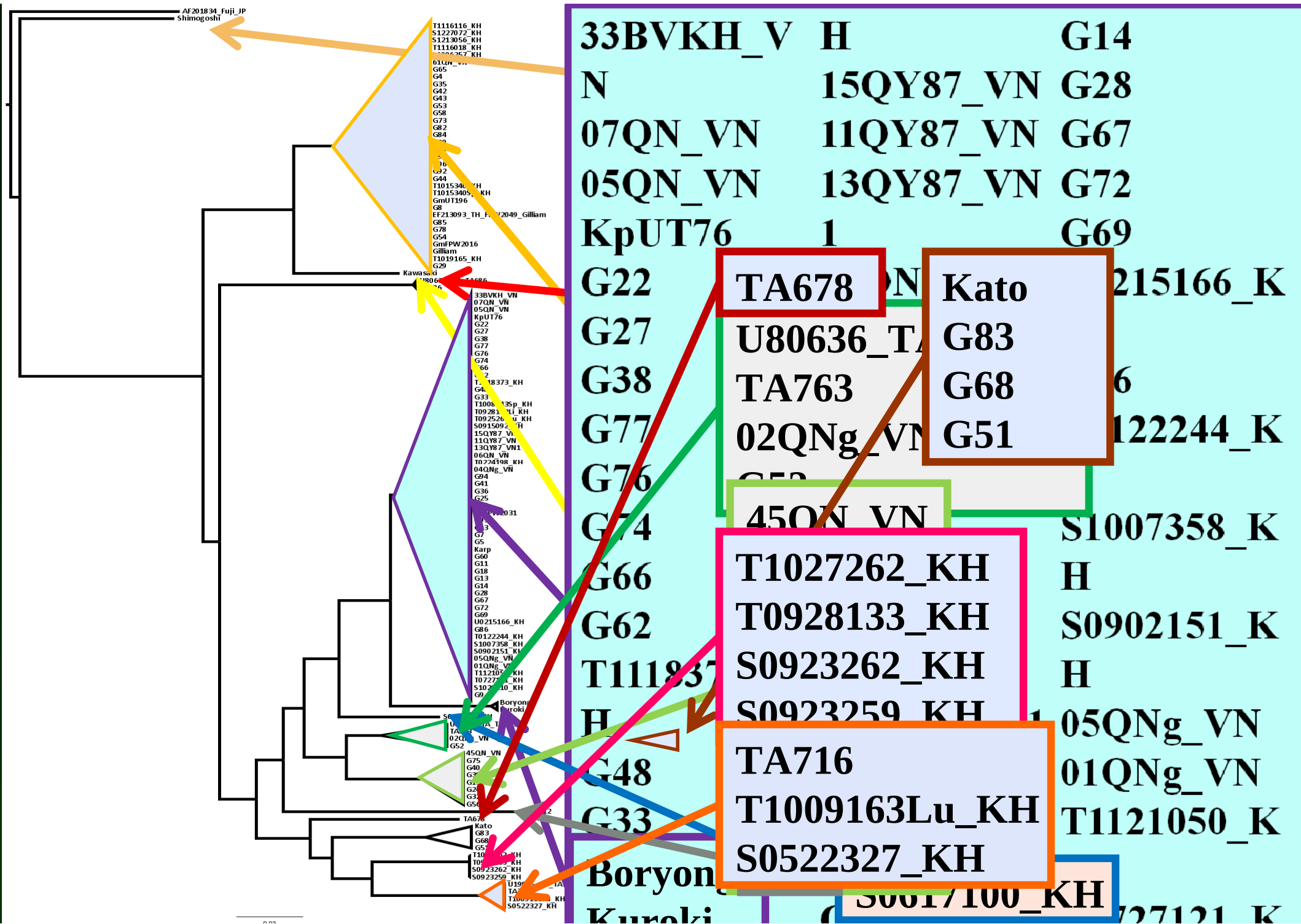
Cambodia

12 provinces
28 strains

Vietnam

3 provinces
12 strains





Conclusion

No geographic clustering

56-kDa

MLST

Distribution depends on small mammals distribution

Distribution is impacted by human activities

Deforestation, land use

Urban growth

Global changes

Distribution most likely to evolve in space and time

Under diagnosed in Cambodia and Vietnam

Clinical samples are not representative (rare and unique events)

Focus on mites and rodents for actual distribution

Priority probably not on epidemiology

Priority probably on awareness and efficient early diagnostic and treatment

Partnership

I.P. Cambodia

V. Duong
K. Blasdel
P. Buchy

CNRS / UM2

L. Gavotte

Calmette Hospital

C. Ra

Mahidol- Siriraj

P. Wongprompitak
W. Anukool
Y. Suputtamongkol
E. Wongsawat
S. Silpasakorn
P. Ekpo

I.P. Nha Trang

Trinh Thi Xuan Mai
Le Viet Lo
Bui Trong Chien

Kantha Bopha Hospital

D. Laurent

Cirad

R. Frutos

Thank you for your attention