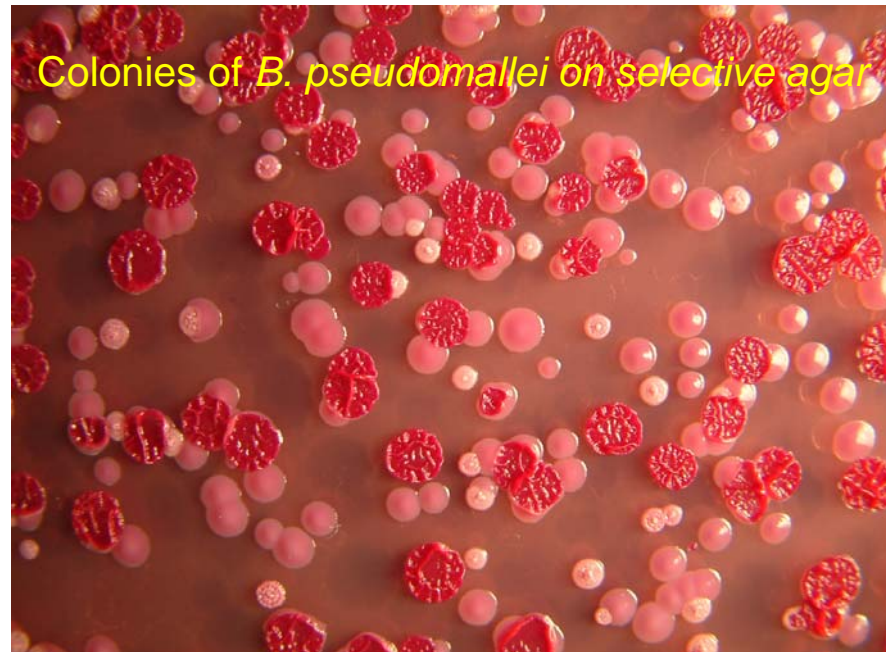


Genetic diversity of *Burkholderia pseudomallei* isolated from agricultural land in northeast Thailand



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Introduction

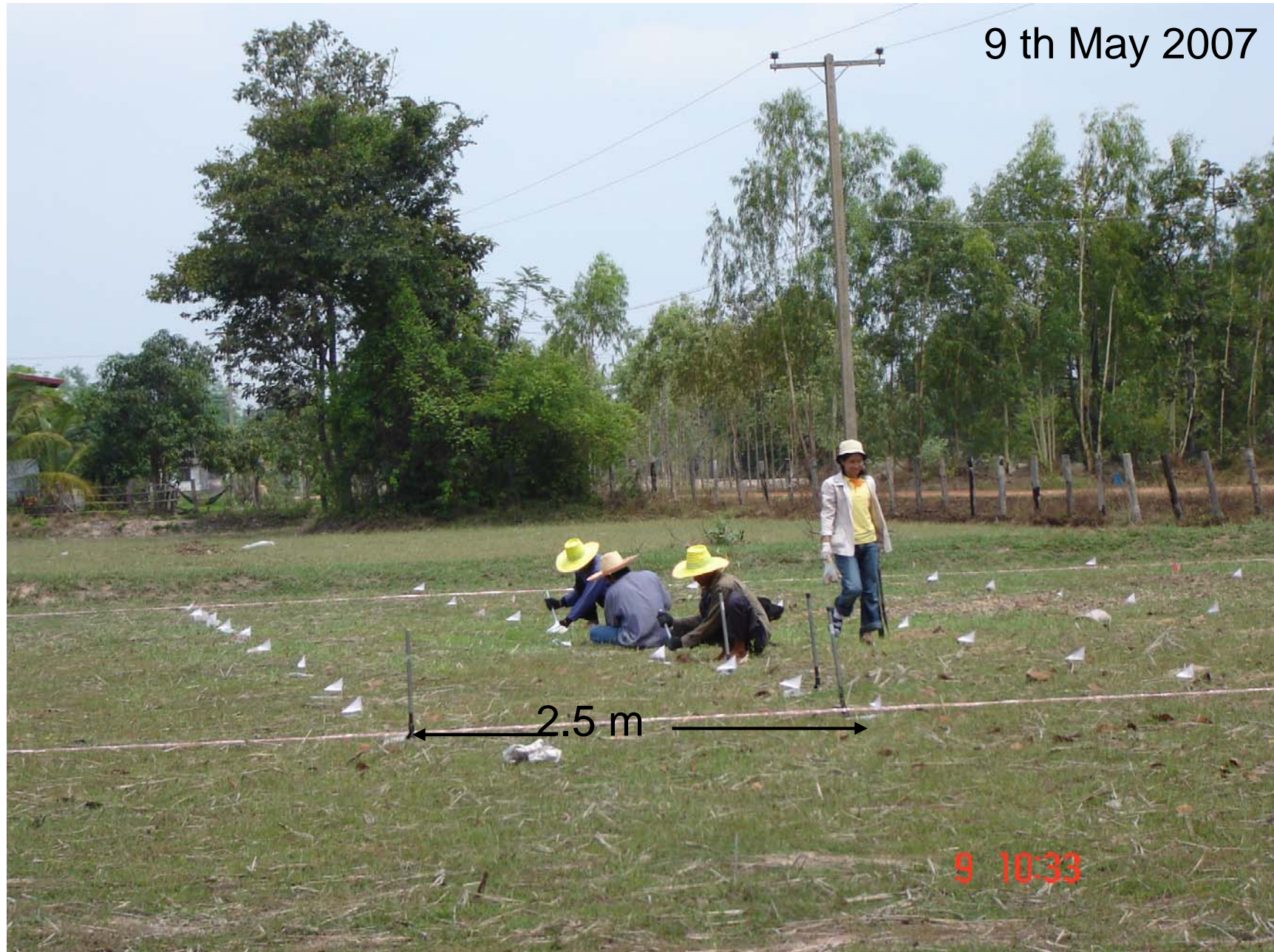


- The soil dwelling Gram-negative bacterium *Burkholderia pseudomallei* is the cause of melioidosis, a serious human infection with a mortality rate of 50% in Thailand and 20% in Australia
- Infection is acquired following bacterial inoculation and contamination of wounds, or more rarely via inhalation or ingestion
- The genetic diversity of this organism in the environment is poorly defined

Objectives

1. Undertake soil sampling in a rice field, where melioidosis is usually acquired
2. Determine the proportion of soil samples positive for *B. pseudomallei* (out of 100 samples)
3. Determine the *B. pseudomallei* load (cfu/gm soil) at each positive site
4. Characterise the colonial appearance (type) on Ashdowns agar
5. Undertake genotyping to determine the genetic diversity of *B. pseudomallei* in soil

Study site: Amphoe Lao Sua Kok, Ubon Ratchathani, northeast Thailand



Overview of methodology

Soil culture for the presence of *B. pseudomallei*



Characterisation of colony morphotype
using an existing algorithm



Genotyping using a combination of PFGE and MLST

Soil culture

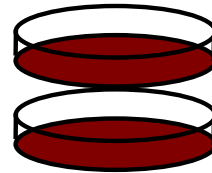
100 g of soil mixed with 100 ml sterile water and left overnight at room temperature



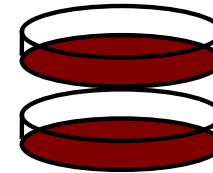
Separate supernatant



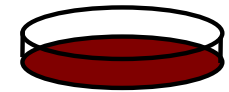
Spread onto 5 plates of Ashdown's selective agar (ASH)



10 µl



100 µl



500 µl

1 ml of supernatant add in to 9 ml of selective enrichment broth



Incubated at 40 °C, 48 h

10 ul of surface liquid plated onto ASH



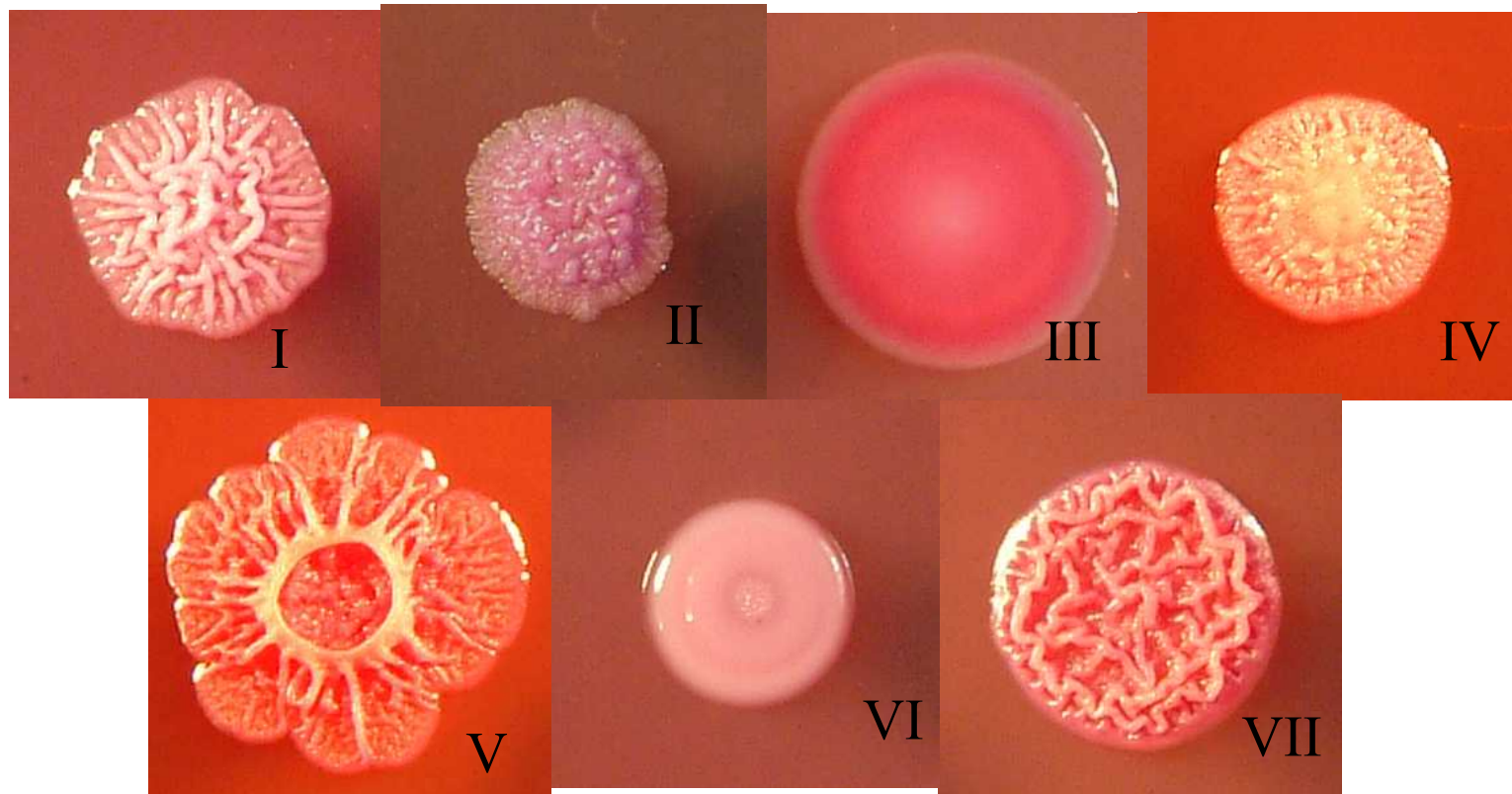
Incubated at 40 °C, 4 days

Incubated at 40 °C, 4 days

Identify and characterise colony morphotype

Characterisation of colony morphotype

Seven morphotypes characterised to date



Genotyping of *B. pseudomallei*

~600 primary colonies were picked from ASH for genotyping by:

- Pulsed field gel electrophoresis (PFGE)

(Analysed using the BioNumerics software version 2.5 (Applied Maths, Belgium))

- Multi Locus Sequence Typing (MLST)



<http://bpseudomallei.mlst.net/>

Pulsed Field Gel Electrophoresis

PFGE is a technique used to separate especially long strands of DNA by length in order to tell differences among samples. It operates by alternating electric fields to run DNA

Multilocus Sequence Typing (MLST)

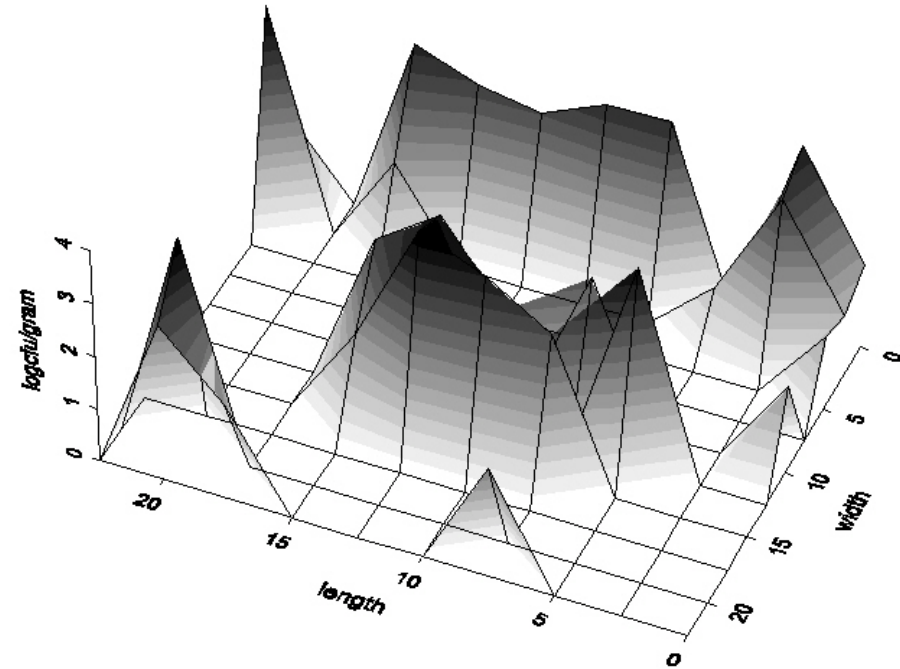
- MLST is a nucleotide sequence based approach for the unambiguous characterization of isolates of bacteria and other organisms via the internet.
- Developed by Godoy et al., 2003 for typing of *B. pseudomallei* and closely related species
- 7 housekeeping genes (*ace*, *lepA*, *lipA*, *nark*, *ndh*, *gltB* and *gmhD*) were designed base on gene sequence of *B. pseudomallei* K96243

Results of soil culture for *B. pseudomallei*

28/100 soil samples were culture positive

Presence & count (log cfu/gm) B. pseudomallei
 median 700 range 3 to >10,000
 IQR 50 to 2810

	1	2	3	4	5	6	7	8	9	10
A	A1		A3	A4	A5	A6	A7		A9	A10
B	B1		B3					B8	B9	B10
C	C1									
D										
E							E7			E10
F				F4	F5	F6		F8		
G				G4	G5	G6	G7			
H										
I		I2								
J		J2	J3				J7			



Colony morphology

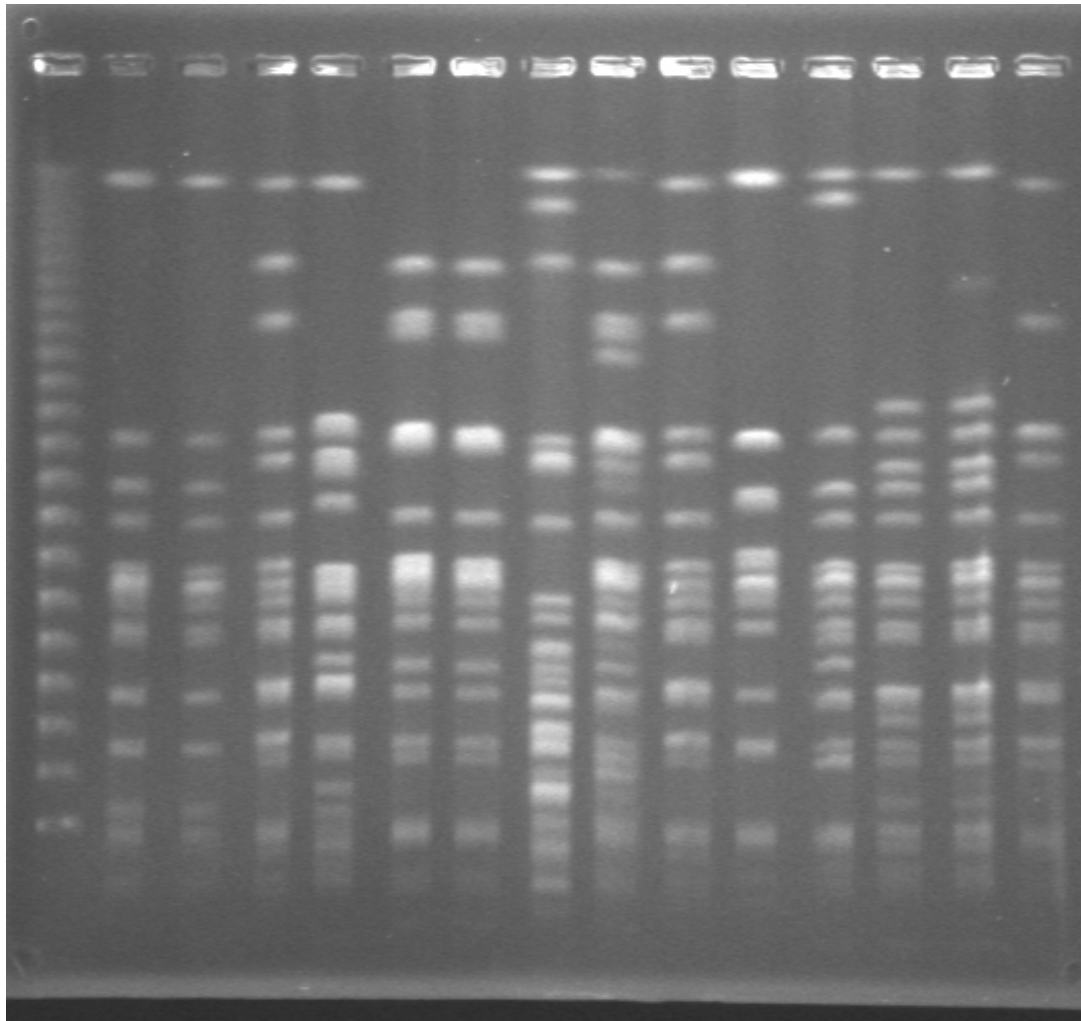
Morphotype I



Only type I was identified
It is the preferential morphotype in the environment

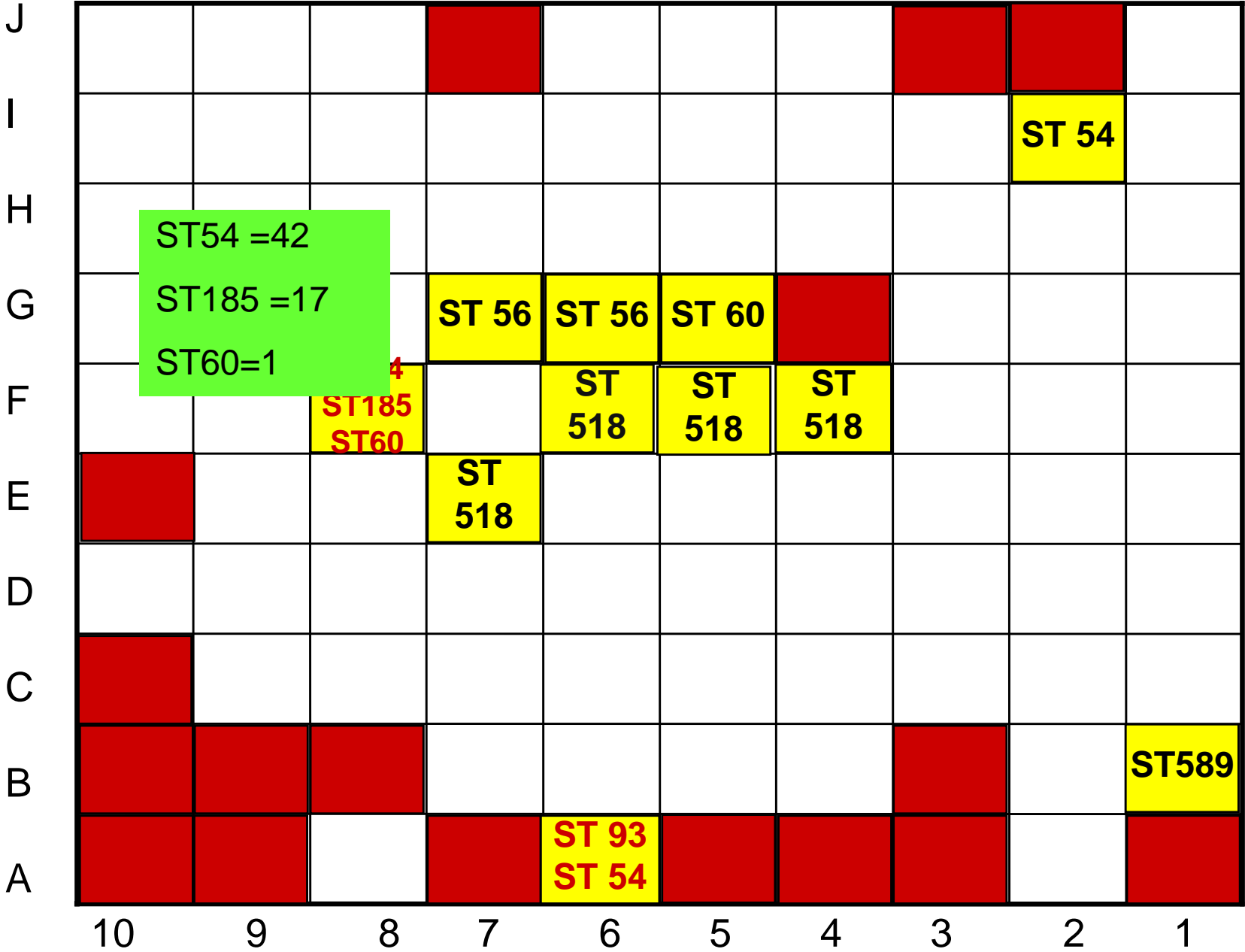
Genotyping using PFGE and MLST

A total of 630 colonies were typing using PFGE. This gave 11 PFGE banding pattern types. MLST of a representative of each type resolved 7 sequence types



PFGE Type	ST
A	ST 93
B, F, K	ST 54
C	ST 589
D, E	ST 518
G	ST 185
H	ST 60
I, J	ST 56

Measures of genetic diversity in 11 positive sites



Summary

- *B. pseudomallei* was present in 28 / 100 sampling points
- Only colony morphotype I was identified in this rice field
- Eleven PFGE types and 7 multilocus sequence type were identified from a total of 11 independent sampling points
- Multiple *B. pseudomallei* genotypes were present within a single soil sample
- Different *B. pseudomallei* genotypes were present at independent but nearby sampling points

Acknowledgements

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- During continuous field electrophoresis, DNA above 30-50 kb migrates with the same mobility regardless of size. This is seen in a gel as a single large diffuse band. If, however, the DNA is forced to change direction during electrophoresis, different sized fragments within this diffuse band begin to separate from each other.
- With each reorientation of the electric field relative to the gel, smaller sized DNA will begin moving in the new direction more quickly than the larger DNA. Thus, the larger DNA lags behind, providing a separation from the smaller DNA.

Table 5. Relationship between 11 PFGE banding pattern and 7 MLST analysis of soil isolates

PFGE Type	ST	MLST profile						
		<i>ace</i>	<i>gltB</i>	<i>gmhD</i>	<i>lepA</i>	<i>lipA</i>	<i>narK</i>	<i>ndh</i>
A	ST 93	1	1	2	1	1	4	1
B,F,K	ST 54	3	1	3	3	1	2	1
C	ST 589	3	1	11	1	1	1	1
D, E	ST 518	1	1	13	2	1	1	1
G	ST 185	1	4	2	2	1	4	1
H	ST 60	3	1	12	1	1	3	1
I, J	ST 56	3	1	4	1	1	4	1

Table 6. Genotyping results for 630 colonies of *B. pseudomallei* from each of independent sampling

Sequence type	Number of colonies per point										
	A6	B1	E7	F4	F5	F6	F8	G5	G6	G7	I2
ST 93	56										
ST 54	1						42				60
ST 589		60									
ST 518			45	60	60	45					
ST 185							17				
ST 60							1	60			
ST 56									60	60	

Result

1. Soil culture

	Number of positive culture points	Number of negative culture points	Total
Disused land	77	23	100
Rice paddy	28	72	100

A possible explanation for the lower number of positive samples in the rice paddy is the detrimental

- Effect of chemical pesticides and fertilisers.
- Difference in positivity between different sampling sites represents natural variation within the region.