# FISH AS THE NATURAL SECOND INTERMEDIATE HOST OF GNATHOSTOMA SPINIGERUM

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**Abstract.** Gnathostomiasis is a helminthic disease most frequently occurring in Thailand. Human infections are usually found to be caused by *Gnathostoma spinigerum*, although five species of the genus *Gnathostoma* exist in Thailand, and three of these are capable of infecting man. In Thailand, 47 species of vertebrates – fish (19), frogs (2), reptiles (11), birds (11) and mammals (4) – have been reported to serve naturally as the second intermediate (and/or paratenic) hosts of *G. spinigerum*. Of these, fish, especially swamp eels (*Monopterus albus*), were found to be the best second intermediate/paratenic hosts: they had the highest prevalence rate and the heaviest infection intensity. However, the scientific names of these fish have been revised from time to time. Therefore, for clarity and consistency, we have summarized the current scientific names of these 19 species of fish, together with their illustrations. We describe one additional fish species, *Systomus orphoides* (*Puntius orphoides*), which is first recorded as a naturally infected second intermediate host of *G. spinigerum*.

#### INTRODUCTION

Several helminthic zoonoses can be transmitted to humans via both marine and freshwater fish. These include capillariasis (caused primarily by Capillaria phillipinensis), gnathostomiasis (Gnathostoma spinigerum), anisakiasis (Anisakis simplex), dioctophymiasis (Dioctophyme renale), eustrongylidiasis (Eustrongylides spp), clonorchiasis (Clonorchis sinensis), opisthorchiasis (Opisthorchis viverrini), echinostomiasis (Echinostoma spp), heterophyiasis (Heterophyes heterophyes), metagonimiasis (Metagonimus *yokogawai*), nanophyetiasis (*Nanophyetus salmincola*) and Diphyllobothriasis (Diphyllobothrium latum) (Beaver et al, 1984; Eastburn et al, 1987; Eberhard et al, 1989: Harrell and Deardorff, 1990: Miyazaki, 1991: Anderson, 1992; Narr et al, 1996). In Thailand, fishborne helminthiases are rather common: the two most prevalent infections among Thai people are opisthorchiasis and gnathostomiasis (Daengsvang, 1980, 1986; Viyanant, 1981; Jongsuksuntigul and Imsomboon, 1998).

Human gnathostomiasis is a disease primarily caused by larval and immature stages of *G. spinigerum* (Daengsvang, 1980; Miyazaki, 1991). However, four other species – *G. hispidum*, *G. doloresi*, *G. nipponicum* and *G. binucleatum* – are also known to

Vol 33 (Suppl 3) 2002

cause disease (Araki, 1986; Ogata *et al*, 1988; Ando *et al*, 1988; Nawa *et al*, 1989; Almeyda-Artigas, 1991; Akahane *et al*, 1998; Almeyda-Artigas *et al*, 2000). There have been at least five species of *Gnathostoma* documented in Thailand: *G. spinigerum*, *G. hispidum*, *G. doloresi*, *G. vietnamicum* and *G. malaysiae* (Dissamarn *et al*, 1966; Daengsvang, 1973, 1980; Kamiya *et al*, 1987); however, only *G. spinigerum* is known to be responsible for human infection in the country (Daengsvang, 1980, 1986; Radomyos and Daengsvang, 1987).

In Thailand, 47 species of vertebrates – fish (19), frogs (2), reptiles (11), birds (11) and mammals (4) – are reported to serve naturally as the second intermediate (and/or paratenic) hosts of *G. spinigerum* (Daengsvang, 1980; Rojekittikhun *et al*, 1989a, 1989b). Of these animals, the fish, especially swamp eels (*Monopterus albus*), have been found to be the best second intermediate/paratenic hosts of the worm: they have the highest prevalence rate and the heaviest infection intensity; moreover, they have been found to harbor at least four species of *Gnathostoma* (Daengsvang, 1980; Rojekittikhun *et al*, 1989a, 1998a, 1998b; Setasuban *et al*, 1991; Akahane *et al*, 1995; Nuamtanong *et al*, 1998).

The Department of Helminthology, Faculty of Tropical Medicine, Mahidol University, has conducted a considerable amount of research into *Gnathostoma* and gnathostomiasis. Advanced third-stage larvae from swamp eels and many other freshwater fish are collected regularly. However, the scientific names of these fish have been subject to periodic revision. In the interest of clarity and consistency, we have summarized the current scientific names of these 19

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species of fish with their illustrations. We also describe one additional fish species, *Systomus orphoides* (*Puntius orphoides*), which is first recorded as a naturally infected second intermediate host for *G. spinigerum*.

## MATERIALS AND METHODS

#### Fish

Fish and eels that had grown naturally in endemic areas were the main target of this study. Fish, dead or alive, were usually purchased from local markets in Nakhon Nayok and Prachin Buri; occasionally fish were obtained from several other provinces beyond Central Thailand. The fish were transported to the laboratory in Bangkok. They were measured, weighed, and identified by species.

#### Press preparation (compression) method

The visceral organs of the big fish and the eels were taken out; the liver was cut into small pieces which were firmly pressed between two thick glass plates and then examined under a dissecting microscope or a big hand lens for the presence of gnathostome larvae. All muscles were cut and scraped out of bones and scales (or skins) and examined using the method used for liver tissue. For tiny fish, the whole body was pressed and examined as described above. The collected larvae were cleaned, counted and identified.

#### **Digestion technique**

The livers or muscles of the fish were chopped and put into a container containing artificial gastric juice (1% HCl and 1% pepsin). They were then incubated for 1-3 hours in a water bath at 37 °C with frequent stirring. The digested tissues were washed several times with normal saline by a simple sedimentation technique. The clear sediments were examined under a dissecting microscope.

#### RESULTS

The 20 species of fish in Thailand that have been found to be naturally infected with *G. spinigerum* larvae are shown in Table 1. Their previous and current scientific names, common names and Thai names together with the maximum number of larvae per fish are also tabulated. Of note is the fact that of the 20 species of 16 genera, 9 genera and 6 species have been changed. Five species, *Boesemania microlepis*, *Chitala ornata, Mastacembelus armatus, Ompok krattensis* and *Systomus orphoides*, harbored only one larva each. Two species, *Micronema apogon* and *Trichogaster trichopterus*, had no record of the maximum number of infected larvae. Swamp eels (*Monopterus albus*) were the most resilient carrier: 2,582 gnathostome larvae were found in the body of one eel. *Systomus orphoides*, the red-cheek barb, was for the first time, found to be a natural second intermediate host of *G. spinigerum*.

Fig 1 shows photographs of the 20 fish with their current scientific and Thai names. Eighteen pictures were taken from fresh specimens, pictures 1.2 and 1.14 were from an illustrated poster (Vidthayanon C. Fishes of Chao Phraya River), and picture 1.16 was drawn by Luang Masya (Smith, 1965). The size of each fish is indicated by a ruler in each picture.

#### DISCUSSION

Almost all of the 20 fish naturally infected with *G. spinigerum* larvae feed on both live or decaying animals and plants. The gourami (*Trichogaster* spp) feeds mainly on tiny living organisms and insects. Therefore, they act more likely as the second intermediate host of the nematode. Eels (*Monopterus* and *Ophisternon*), snake-head fish (*Channa* spp) and catfish (*Clarias* spp) are carnivorous creatures. Not only do they act as second intermediate hosts, they also serve as paratenic hosts. This, without doubt, increases their chance of becoming more and heavily infected.

The drum fish (Boesemania microlepis) is a marine fish; it can, however, live in fresh water for periods of time. Kasemsuthi et al (1983) have demonstrated how this kind of fish was infected: Java tilapias (Oreochromis mossambica, previously Tilapia mossambica), an amphidromous fish, was collected from the sea at Bang Saen Beach and then orally infected with G. spinigerum larvae after the salinity of the water was reduced to zero. The salinity of the water was then gradually increased to 30 ppt and the fish were examined at days 20 and 40 post-infection. The infection rates were 71.4% and 58.8%, respectively; the corresponding survival rates of the recovered larvae were 87.5% and 52.6%, respectively. The flesh of these infected fish, containing G. spinigerum larvae, was fed to a real marine fish, Epinephelus spp, which was examined about 1-2 months later: it was infected and the larvae were still alive (Kasemsuthi, personal communication). Another experiment done in 1974 by Nithi-Uthai showed that two species of brackish water fish, Mugil sp and Chanos chanos, could be infected with G. spinigerum larvae by both forced- and self-feeding. The percentages of infection were 56.0% and 80.0%, respectively, for forced feeding, and 53.3% and 30.0%, respectively, for self-feeding. However, examination of four species of brackish water fish and

Table 1 The twenty species of fish in Thailand found to be naturally infected with <i>Gnathostoma spinigerum</i> larvae.
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Curren	Current scientific name	Previous name	Common name	Thai name	Length (cm) <sup>a</sup>	Max. no. of larvae/fish
Ι.	Anabas testudineus	Same	Common climbing perch	Pla mo Thai	10-23	14
5.	Arius caelatus	Tachysurus caelatus	Engraved catfish	Pla kot	20-29	5
З.	Boesemania microlepis	Nibea soldado	River drumfish	Pla ma	17-60	1
4	Channa striata	Ophicephalus striatus	Striped snake-head fish	Pla chon	30-70	153
5.	Channa micropeltes	<b>Ophicephalus micropeltis</b>	Giant snake-head fish	Pla chado	30-75	12
6.	Channa lucius	<b>Ophicephalus lucius</b>	Blotched snake-head fish	Pla krasong	20-65	12
7.	Clarias batrachus	Same	Batrachian walking catfish	Pla duk dan	16-40	9
8.	Clarias macrocephalus	Same	Gunther's walking catfish	Pla duk ui	15-35	16
9.	Chitala ornata	Notopterus chitala	Spotted featherback	Pla krai	35-85	1
10.	Kryptopterus cryptopterus	Same	ż	Pla kha kai	11-16	5
11.	Macrognathus siamensis	Same	Spotted spiny eel	Pla lot chut	15-30	8
12.	Mastacembelus armatus	Same	Armed spiny eel	Pla krathing dam	20-70	1
13.	Micronema apogon	Kryptopterus apogon	Common sheatfish	Pla nam ngoen	15-77	ż
14.	Monopterus albus	Fluta alba	Swamp eel	Pla lai na	30-95	2,582
15.	Ompok krattensis	Ompok bimaculatus	Sheatfish	Pla nua on	15-20	1
16.	Ophisternon bengalense	Synbranchus bengalensis	Eel-like fish, Bengal eel	Pla lat, pla lai	25-53	40
17.	Systomus orphoides	Puntius orphoides	Red-cheek barb	Pla kaem cham	8-22	1
18.	Trichogaster pectoralis	Same	Snake skin gourami	Pla salit	15-20	5
19.	Trichogaster microlepis	Same	Moonlight gourami	Pla kradi nang	6-15	5
20.	Trichogaster trichopterus	Trichopodus trichopterus	Three-spot gourami	Pla kradi mo	5-12	ż

<sup>a</sup> Normal length of common fish (Smith, 1965; Department of Fisheries, Ministry of Agriculture and Cooperatives, 1997).



Fig 1- Illustrations of the 20 fish species in Thailand found to be naturally infected with *G. spinigerum* larvae. Their current scientific names, with Thai names in parentheses, are as follows: 1.1. *Anabas testudineus* (pla mo Thai), 1.2. *Arius caelatus* (pla kot), 1.3. *Boesemania microlepis* (pla ma), 1.4. *Channa striata* (pla chon), 1.5. *Channa micropeltes* (pla chado), 1.6. *Channa lucius* (pla krasong), 1.7. *Clarias batrachus* (pla duk dan), 1.8. *Clarias macrocephalus* (pla duk ui), 1.9. *Chitala ornata* (pla krai), 1.10. *Kryptopterus cryptopterus* (pla kha kai), 1.11. *Macrognathus siamensis* (pla lot chut), 1.12.

#### FISH AS HOSTS OF G. SPINIGERUM



Mastacembelus armatus (pla krathing dam), 1.13. Micronema apogon (pla nam ngoen), 1.14. Monopterus albus (pla lai na), 1.15. Ompok krattensis (pla nua on, pla cha-on), 1.16. Ophisternon bengalense (pla lat, pla lai), 1.17. Systomus orphoides (pla kaem cham), 1.18. Trichogaster pectoralis (pla salit), 1.19. Trichogaster microlepis (pla kradi nang), 1.20. Trichogaster trichopterus (pla kradi mo).

34 species of marine fish from the Gulf of Thailand and adjacent areas for natural infection of gnathostome larvae did not demonstrate any infection (Nithi-Uthai, 1974).

In Thailand, there are about 145 species of cyprinoid fish (Smith, 1965). Until this study, none had been found to serve as the host of *Gnathostoma*. In this study, the red-cheek barb (*Systomus orphoides*) was shown for the first time to be infected. If a great number of fish of any species were to be subjected to careful examination, it would be more than likely that they too would find their way onto the list of naturally infected second intermediate hosts of *Gnathostoma*.

### ACKNOWLEDGEMENTS

The authors would like to thank Dr Chavalit Vidthayanon of the Museum Division, Department of Fisheries, for identifying the fish.

#### REFERENCES

- Akahane H, Setasuban P, Nuamtanong S, *et al.* A new type of advanced third-stage larvae of the genus *Gnathostoma* in freshwater eels, *Fluta alba*, from Nakhon Nayok, central Thailand. *Southeast Asian J Trop Med Public Health* 1995;26:743-7.
- Akahane H, Sano M, Kobayashi M. Three cases of human gnathostomiasis caused by *Gnathostoma hispidum*, with particular reference to the identification of parasitic larvae. *Southeast Asian J Trop Med Public Health* 1998;29:611-4.
- Almeyda-Artigas RJ. Finding of *Gnathostoma binucleatum* n. sp. (Nematoda: Spirurida) in wild felids and the role of fresh-water and oligohaline fishes as human gnathostomiasis vectors in the lower basin of Papaloapan river, Oaxaca-Veracruz, Mexico. An Inst Cienc Mar Limnol Univ Nal Autón México 1991;18:137-55 (Spanish with English abstract).
- Almeyda-Artigas RJ, Bargues MD, Mas-Coma S. ITS-2 rDNA sequencing of *Gnathostoma* species (Nematoda) and elucidation of the species causing human gnathostomiasis in the Americas. *J Parasitol* 2000;86:537-44.
- Anderson RC. Nematode parasites of vertebrates: their development and transmission. Cambridge: CAB International, University Press, 1992.
- Ando K, Tanaka H, Taniguchi Y, *et al.* Two human cases of gnathostomiasis and discovery of a second intermediate host *of Gnathostoma nipponicum* in Japan. *J Parasitol* 1988;74:623-7.

- Araki T. Gnathostomiasis Parasitic disease caused by eating raw loaches. *Kansen Ensyou Meneki* 1986;16:110-1 (in Japanese).
- Beaver PC, Jung RC, Cupp EW. Clinical parasitology. 9<sup>th</sup> ed. Philadelphia: Lea & Febiger, 1984.
- Daengsvang S. First report on Gnathostoma vietnamicum Le-Van-Hoa, 1965 from urinary system of otters (Aonyx cinerea, Illiger) in Thailand. Southeast Asian J Trop Med Public Health 1973;4:63-70.
- Daengsvang S. A monograph on the genus Gnathostoma and gnathostomiasis in Thailand. Tokyo: Southeast Asian Medical Information Center (SEAMIC), International Medical Foundation of Japan, 1980.
- Daengsvang S. Gnathostoma spinigerum and human gnathostomiasis. A review. In: Sucharit S, et al, eds. The 25<sup>th</sup> Anniversary of the Faculty of Tropical Medicine, Mahidol University. Bangkok: Krung Siam Press, 1986:124-47.
- Department of Fisheries, Ministry of Agriculture and Cooperatives. Illustrated fish and aquatic animals of Thailand. Bangkok: Suksaphanphanit 1997 (in Thai).
- Dissamarn R, Thirapat K, Aranyakanada P, *et al.* Studies on morphology and life history of *G. doloresi* and *G. hispidum* in Thailand. *J Thai Vet Med Assoc* 1966;17:1.
- Eastburn RL, Fritsche TR, Terhune CA Jr. Human intestinal infection with *Nanophyetus salmincola* from salmonid fishes. *Am J Trop Med Hyg* 1987;36:586-91.
- Eberhard ML, Hurwitz H, Sun AM, *et al.* Intestinal perforation caused by larval *Eustrongylides* (Nematoda: Dioctophymatoidae) in New Jersey. *Am J Trop Med Hyg* 1989;40:648-50.
- Harrell LW, Deardorff TL. Human nanophyetiasis: transmission by handling naturally infected coho salmon (*Oncorhynchus kisutch*). J Infect Dis 1990;161:146-8.
- Jongsuksuntigul P, Imsomboon T. Epidemiology of opisthorchiasis and national control program in Thailand. *Southeast Asian J Trop Med Public Health* 1998;29:327-32.
- Kamiya H, Kamiya M, Ohbayashi M, et al. Gnathostoma malaysiae Miyazaki and Dunn, 1965 from Rattus surifer in Thailand. Southeast Asian J Trop Med Public Health 1987;18:121-6.
- Kasemsuthi R, Panassumpon P, Siriratanachai S.

Transmission of *Gnathostoma spinigerum* in amphidromous fish (*Tilapia mossambica*). J Aquat Anim Dis 1983;6:1-7.

- Miyazaki I. An illustrated book of helminthic zoonosis. Tokyo: International Medical Foundation of Japan, 1991.
- Narr LL, O' Donnell JG, Libster B, *et al.* Eustrongylidiasis - a parasitic infection acquired by eating live minnows. *J Am Osteopath Assoc* 1996;96:400-2.
- Nawa Y, Imai J, Ogata K, et al. The first record of confirmed human case of *Gnathostoma doloresi* infection. J Parasitol 1989;75:166-9.
- Nithi-Uthai S. Studies on *Gnathostoma spinigerum*. I. Development of eggs, and larvae in cyclops. II. Experimental and natural infections in brackish water fishes. Bangkok: Mahidol Univ 1974. MSc thesis.
- Nuamtanong S, Waikagul J, Anantaphruti MT. Gnathostome infection in swamp eels, *Fluta alba*, in central Thailand. *Southeast Asian J Trop Med Public Health* 1998;29:144-7.
- Ogata K, Imai J, Nawa Y. Three confirmed and five suspected human cases of *Gnathostoma doloresi* infection found in Miyazaki Prefecture, Kyushu. *Jpn J Parasitol* 1988;37:358-64.
- Radomyos P, Daengsvang S. A brief report on *Gnathostoma spinigerum* specimens obtained from human cases. *Southeast Asian J Trop Med Public Health* 1987; 8:215-7.

- Rojekittikhun W, Pubampen S, Hiranyachattada P, et al. A survey on the infective larvae of Gnathostoma spinigerum in fresh water fish sold in the local markets of Bangkok. J Trop Med Parasitol 1989a;12:7-12 (Thai with English abstract).
- Rojekittikhun W, Daengswang S, Pubampen S, *et al.* A survey on the infective larvae of *Gnathostoma spinigerum* in rats at Nakhon Nayok and Prachin Buri. *J Trop Med Parasitol* 1989b;12:13-6 (Thai with English abstract).
- Rojekittikhun W, Pubampen S, Waikagul J. Swamp eels (*Fluta alba*), the genuine second intermediate host of *Gnathostoma* in Thailand. *J Trop Med Parasitol* 1998a;21:44-5.
- Rojekittikhun W, Pubampen S, Waikagul J. Seasonal variation in the intensity of *Gnathostoma* larvae in swamp eels (*Fluta alba*) sold in a local market of Bangkok. *Southeast Asian J Trop Med Public Health* 1998b;29:148-53.
- Setasuban P, Nuamtanong S, Rojanakittikoon V, et al. Gnathostomiasis in Thailand: a survey on intermediate hosts of *Gnathostoma* spp with special reference to a new type of larvae found in *Fluta alba. Southeast Asian J Trop Med Public Health* 1991; 22 (suppl): 220-4.
- Smith HM. The fresh-water fishes of Siam, or Thailand. New Jersey: TFH Publications, 1965.
- Viyanant V. Opisthorchiasis in Thailand: a review. J Parasitol Trop Med Assoc Thai 1981;4:87-106.