PROMOTION OF AFLATOXIN B₁- INDUCED HEPATOCARCINOGENESIS BY DICHLORODIPHENYL TRICHLOROETHANE (DDT)

Subhkij Angsubhakorn¹, Apichat Pradermwong¹, Kanthimani Phanwichien² and Sudarat Nguansangiam³

¹Department of Pathobiology, Faculty of Science, Mahidol University, Bangkok; ²Department of Zoology, Faculty of Science, Kasetsart University, Bangkok; ³Department of Pathology, Bangkok Metropolitan Administration Medical College and Vajira Hospital, Bangkok, Thailand

Abstract. A study of the effect in rats of dichlorodiphenyl trichloroethane (DDT) on hepatocarcinogenesis that is initated by aflatoxin B_1 (AFB₁). In the first experiment, Buffalo rats were given a single oral dose of AFB₁ (5 mg/kg) followed by dietary DDT (100 ppm) for 20 weeks. Neoplastic nodules were observed in 1 of the 14 AFB₁-exposed rats, compared with 3 of the 19 rats in the AFB₁/DDT group. In the second experiment, Wistar rats were given dietary aflatoxin B_1 (4 ppm) for 6 weeks followed by a 6-week exposure to DDT (500 ppm) in a plain semisynthetic diet. Five altered hepatic foci were displayed by seven rats in the AFB₁ group, compared with 6 foci and one neoplastic focus in five of the AFB₁/DDT rats at 32 weeks. Subsequently, the AFB₁ group produced 8 (27.5%) tumor-bearing rats while 10 of the 28 (35.7%) AFB₁/DDT-exposed rats were tumor-bearing by 60 weeks. The results suggest that DDT slightly potentiates hepatocarcinogenesis induced by either a single dose of AFB₁ or short term-dietary AFB₁.

INTRODUCTION

Aflatoxin B_1 (AFB₁), a potent, naturally occurring hepatocarcinogen produced by Aspergillus flavus and A. parasiticus, has attracted wide interest because of its association with a high incidence of human liver cancer, particularly in Southeast Asia (Shank et al, 1972 a,b,c; Angsubhakorn, 1983) and Africa (Alpert et al, 1971; Peers and Linsell, 1973; Peers et al, 1976; Van Rensburg et al, 1985). Furthermore, in an analysis involving 10 of the smaller subregions of Swaziland, aflatoxin exposure emerged as a more important correlate of variation in liver cancer incidence than the prevalence of hepatitis B virus infection (Peers et al, 1987; Montesano et al, 1997). The problems of aflatoxins in foods and feedstuffs

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in Thailand need attention, on both economic and health grounds. Dichlorodiphenyl trichloroethane (DDT) is also known to be a potential health hazard due to its chronic ingestion from the environment. The highest levels of fatsoluble organochlorine compounds, such as DDT, are found in the dietary fat of meat, fish and poultry and in diary products; an increase in the intake of animal fat may result in higher dietary exposure to organochlorine compounds (Wissermann et al, 1972; Gorchev and Jelinik, 1985). DDT has been widely used in Thailand since 1950: both in agricultural and public health programs, and especially for malaria eradication (Suwankerd and Prajakwong, 1995), it has been found to be a common contaminant of various food and human milk. DDT is still essential for the killing of malarial mosquitos in many tropical countries and was finally exempted by the Treaty on Persistent Organic Pollutants (POP) by the representatives of 122 countries, at a meeting in Johannesburg in December 2000, because there was no effective substitute for mosquito control. (Kaizer and

Correspondence: Dr Subhkij Angsubhakorn, Department of Pathobiology, Faculty of Science, Mahidol University, Rama 6 Road, Bangkok 10400, Thailand. Tel: 66 (0) 2201-5573; Fax: 66 (0) 2246-1379 E-mail: scsag@mahidol.ac.th

Enserink, 2000).

DDT has been shown in chronic feeding experiments with rats to produce a small number of low-grade hepatic cell carcinomas and nodular adenomatoid hyperplasia (Fitzhuch and Nelson, 1947). Other studies with DDT in rats have provided no evidence of carcinogenicity (Haag *et al*, 1950; Cameron and Cheng, 1951; Treon and Cleveland, 1955; Klimmer, 1955; Klimbrough *et al*, 1964; Deichmann *et al*, 1967). In lifespan studies, the incidence of liver tumors in rats given 500 parts per million (ppm) of technical DDT was 56% and 35% in females and males respectively (Rossi *et al*, 1977).

It has been shown that several epigenetic carcinogens, probably of the promotor class like DDT, enhence the carcinogenic effect of previously administered genotoxic carcinogens in rats such as acetylaminofurene (AAF) and 3'methyl-4-dimethyl-aminoazobenzene (3'Me DAB), which are not carcinogenic by themselves (Peraino et al, 1975; Kitagawa et al, 1984). How epigenetic hepatocarcinogens work is being clarified by recent advances in our understanding of the mechanism of liver tumor promotion. Our studies were initiated in order to investigate the carcinogen enhancing effect of DDT on AFB, that had been administered previously as either a single oral dose or a short term dietary regimen.

MATERIALS AND METHODS

Animals

Male Buffalo rats, from our own colony, aged 6 weeks and weighing about 80 g were used in Experiment 1. In Experiment 2, we used male Wistar rats of at least 1 year of age that weighed 400-500 g; these rats were purchased from the National Laboratory Animal Center (NLAC), Mahidol University, Salaya. On arrival, they were randomly divided into groups of 5 rats per cage, and were adapted to housing conditions for at least 7 days before starting the experiment. All animals were kept in a controlled lighting environment (12 hours dark/light cycle) at a constant temperature of 27°C. Food and water were available *ad libitum* and cared for according to the guidelines of the National Laboratory Animal Center (NLAC).

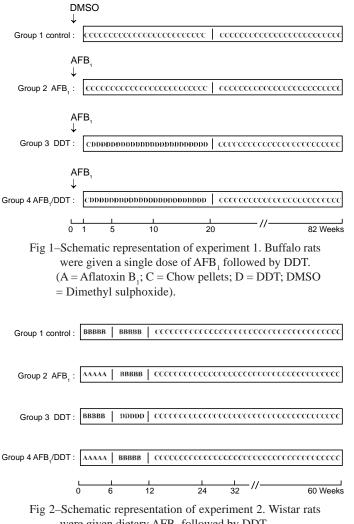
Experiment 1: Powdered Chow pellets (Gold Coin Mills Ltd, Jurong, Singapore) were used as the control diet. DDT (p,p-DDT 86%) was dissolved in 50 ml corn oil and the solution was stirred with chow powder to a final concentration of 100 ppm in a mechanical mixer (Ikeden MS-20, Tokyo Ikeden Co Ltd, Japan).

Experiment 2: The semi-synthetic diet (Angsubhakorn *et al*, 1981) was prepared weekly. AFB_1 (Batch No. 10691, Makor Chemicals Ltd, Jerusalem, Israel) was added to the plain semisynthetic diet after dissolving in a small amount of analytical grade acetone (JT Baker Chemical Co, USA) and corn oil. The acetone was then evaporated to minimize aflatoxin decomposition. DDT was also mixed with corn oil and added to the diet at a final concentration of 100 and 500 ppm.

Experimental design

Experiment 1: The experimental design is outlined in Fig 1. AFB, was dissolved in dimethylsulphoxide (DMSO, BDH, Poole, England) and was immediately administered by intragastric intubation. One hundred Buffalo rats, except for those 27 rats given 0.1 ml DMSO only (group 1), were given AFB₁ at a dose of 5 mg/kg body weight. One week after DMSO or AFB, intragastric intubation, all rats were fed chow pellets (groups 1 and 2) experimental diets (groups 3 and 4) for 20 weeks and then chow pellets until the end of the experiment (84 weeks). Five rats from each group were killed at 1, 5 and 10 weeks after receiving their respective diets. The remaining rats were killed at 82 weeks, when the experiment was terminated.

Experiment 2: In this experiment (Fig 2), 176 male Wistar rats were randomly assigned to 4 treatment groups, as follows. Group 1 (Control): 31 rats were fed plain semisynthetic diet for 12 weeks, followed by chow pellets; Group 2 (AFB₁): 39 rats received a basal diet con-



were given dietary AFB_1 followed by DDT. (A = Aflatoxin B₁; B = Basal diet; C = Chow pellets; D = DDT).

taminated with 4 ppm AFB_1 for 6 weeks, followed by a plain basal diet for 6 weeks, and then chow pellets; Group 3 (DDT): 36 rats were given the basal diet for 6 weeks, followed by DDT for 6 weeks, and then chow pellets; Group 4 (AFB₁/DDT): 53 rats were given dietary AFB₁ for 6 weeks, DDT for 6 weeks, and then chow pellets until the end of the experiment (60 weeks).

At the end of the experiment, all the rats were fasted for 18 hours before being killed. Serum was obtained from clotted whole blood; serum glutamic oxaloacetic transaminase (SGOT) and serum glutamic pyruvic transaminase (SGPT) levels were determined by standard methods (Reitman and Frankel, 1957). The total protein concentrations of the sera were determined by the method of Gornall *et al* (1948) and the serum albumin concentration was determined using Rodkey's method (1965).

Data collection and analysis

A complete necropsy was performed on each rat; each liver was weighed and carefully examined. Tissues were fixed in 10% buffered formalin, embedded in Paraplast-plus (Monoject, St Louis, MO). Microscopic changes in the liver were diagnosed according to previous descriptions (Squire and Levitt, 1975; Steward *et al*, 1980). All results are expressed as group arithmetic mean ± standard deviation. Statistical comparisons of group means were carried out using Student's *t*-test.

RESULTS

Experiment 1

Rats in group 2 (AFB₁), group 3 (DDT), and group 4 (AFB₁/DDT) showed significant decreases in body weight compared with the corresponding controls (p < 0.05) at weeks 1 and 5, weeks 1,5 and 10, and weeks 1,5,10 and 82 respectively (Table 1). The relative liver weights of rats in group 4 (AFB₁/DDT) and group 3 (DDT) showed a significant increase compared with the corresponding control (p < 0.05) in weeks 1,5 and 10, and in week 10 respectively. The DDT intake calculated on the basis of diet consumption for each group showed no significant difference.

During the first 10 weeks, the control rats (group 1) which had been given only a single oral dose of DMSO, did not develop any lesions except oval cell proliferation during the first

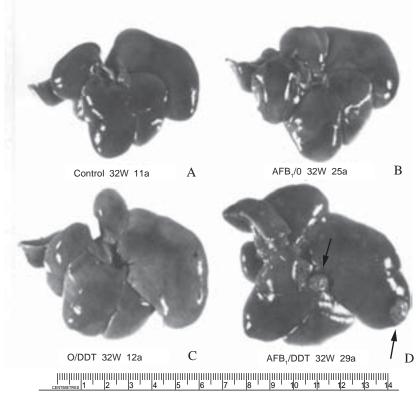


Fig 3–Gross appearance of Wistar rat liver at 32 weeks from 4 groups. A =Control; B=AFB₁; C = DDT; and D= AFB₁/DDT. A, B and C are normal; D has a tumor (arrowed).

week of the experiment. Megalocytosis was found in all 4 rats in group 4 (AFB $_1$ /DDT) by 10 weeks.

At the end of the experiment, 15 of the 19 rats in group 4 (AFB₁/DDT) showed varying degrees of hypertrophic hepatocytes with cytoplasmic inclusions. The inclusion bodies appeared mostly in the AFB₁ pretreated group. A greater number of altered cell foci were found in the livers of rats that had been pretreated with a single dose of AFB₁ followed by DDT. Neoplastic nodules were observed in 1 of the 14 AFB₁-exposed rats, compared with 3 of the 19 rats in the AFB₁/DDT group.

Experiment 2

Average chemical intake, diet consumption, relative liver weight, and mortality of the

rats are summarized in Table 2. The body weights of the rats fed the AFB,/ DDT diet (group 4) had significantly decreased by week 32 when compared with the control rats (group 1) and the rats given DDT alone (group 3) (p < 0.05). The liver weights of the rats in group 4 (AFB,/DDT diet) were significantly greater in comparison with those of the AFB, and the DDT groups by week 32 (p < 0.05). The AFB, intake of the group 2 rats (AFB₁) was slightly higher than in group 4 (AFB,/ DDT).

Serum analysis

The SGOT and SGPT levels were slightly increased in all treated groups at all times when compared with the controls. Total protein and albumin were slightly decreased in most treated

groups when compared with the controls. The albumin of the rats that has been fed the AFB₁ diet decreased significantly to a minimum of 2.3 g/100 ml at 32 weeks (p < 0.05). In contrast, the albumin of the rats that had been fed the DDT diet showed a significant increase: to a maximum of 4.5 g/100 ml by week 60 (p < 0.05).

Liver pathology

At week 32, there were no remarkable changes to the liver surface of the controls (Fig 3). The livers showed a normal surface and were reddish brown. The livers of all treated groups were darkened and had enlarged, slightly blunt edges. Microscopic findings are summarized in Table 4. Hepatic cells in the DDT-treated rats were considerably enlarged, with vacuoliza-

Group	Treatment	No. of rat	DDT intake (mg)	Body weight (g)	Relative liver weight (%)
	1 week				
1	Control	5	-	128 ± 4^{a}	3.2
2	AFB ₁	5	-	99 ± 4 ^b	3.6
3	DDT	5	5.5	122 ± 4 ^b	3.4
4	AFB ₁ /DDT	5	6.1	109 ± 5^{b}	5.5 ^b
	5 weeks				
1	Control	5	-	223 ± 7	2.7
2	AFB ₁	5	-	174 ± 7 ^b	2.6
3	DDT	5	41.7	175 ± 3 ^b	3.4
4	AFB ₁ /DDT	5	39.6	185 ± 4^{b}	4.2 ^b
	10 weeks				
1	Control	5	-	277 ± 5	2.4
2	AFB ₁	5	-	280 ± 13	2.6
3	DDT	5	98.1	261 ± 13^{b}	3.6 ^b
4	AFB ₁ /DDT	5	108.0	245 ± 10^{b}	3.9 ^b
	82 weeks				
1	Control	12	-	382 ± 32	2.3
2	AFB ₁	14	-	360 ± 25	2.5
3	DDT	22	259.8	385 ± 10	2.8
4	AFB ₁ /DDT	19	260.7	377 ± 4 ^b	2.3

Table 1 Group mean body and liver weights and chemical intakes for Buffalo rats fed DDT after a single oral dose of AFB, in experiment 1.

^aMean ± standard deviation,

^bSignificantly different the from control. (p < 0.05)

			Т	abl	e 2				
Mortality	and	weight	gain	of	Wistar	rats	in	experiment	2.

Group	Treatment	No. of rats at start		of rats ificed eek:	Body gain (at wee	0,	-	No. o	of dea	aths a	it wee	ks o	f tre	atme	nt
			32	60	32	60	9	12	15	18	24	30	38	53	58
1	Control	30	7	18	118±35	47±21	-	-	-	1	1	1	1	-	1
2	4 ppm AFB	35	7	25	38±28	15±5	-	-	2	-	-	-	-	1	-
3	500 ppm DDT	35	6	19	73±13	52±23	3	3	1	-	-	1	-	1	1
4	AFB ₁ /DDT	43	5	28	25±15 ^a	33±11	5	3	-	-	-	-	-	-	2

^aStatistically different from the control group (p < 0.05) and statistically different from the DDT contaminated group (p < 0.05).

tion, degeneration, necrosis, varying degrees of change in fat content, and a moderate degree of oval-cell proliferation. All AFB₁-treated rats showed a massive proliferation of oval cells and the development of altered cell foci which were scattered through-

	Table 3			
Average chemical intake, di	et consumption and	relative liv	ver weight o	of Wistar rats in
	experiment	2.		

Group	Treatment	Total ch consumed		Basal diet consumed g/rat/day	Chow pellet consumed g/rat/day -		ve liver ht (%)
		AFB_1	DDT	g/rat/day	g/rat/day	week 32	week 60
1	Control	-	-	14.3	20.7	2.2	2.8
2	4 ppm AFB ₁	2.9±1	-	14.8	19.3	2.7	2.6
3	500 ppm DDT	-	250±1	13.4	20.0	3.5ª	2.6
4	AFB ₁ /DDT	2.6±0.3	171±67	13.2	23.1	3.6 ^{a,b}	2.8

^aStatistically different from control group (p < 0.05) and statistically different from the AFB₁ contaminated group (p < 0.05).

		32 V	Weeks		60 Weeks					
Description	Control (7)	AFB ₁ (7)	DDT (7)	AFB ₁ /DDT (5)	Control (18)	AFB ₁ (29)	DDT (19)	AFB ₁ /DDT (28)		
Portal triads										
Mononuclear-cell infiltration	0	2	1	1	2	0	0	0		
Oval-cell proliferation	0	5ª	7 ^b	5 ^b	0	6ª,8 ^b ,7 ^c	1ª,2 ^b ,1 ^c	1ª,6 ^b ,4 ^c		
Bile-cell proliferation	0	0	0	0	0	4	4	1		
Cyst	0	0	0	0	0	2	0	0		
Kupffer										
Hypertrophy	0	1	5	4	0	1	13	9		
Hyperplasia	0	1	2	1	0	10	11	5		
Hepatocytes										
Enlargement	0	1	7	5	0	13	3	15		
Vacuolization	0	0	5	3	0	7	8	8		
Fatty change	0	0	7	5	1	6	8	11		
Degeneration/necrosis	0	4	7	3	1	14	9	10		
Hyalin/inclusion bodies	0	0	0	1	0	8	11	13		
Altered cell foci										
Acidophilic cell foci	0	1	0	5	0	16	1	15		
Basophilic cell foci	0	0	0	0	0	12	0	8		
Clear cell foci	0	4	0	1	1	1	0	0		
Neoplastic nodules	0	0	0	1	0	9	1	9		
Carcinoma										
Hepatocellular carcinoma (HCC)	0	0	0	0	0	6	0	7		
Cholangiocellular carcinoma (CCC)	0	0	0	0	0	1	0	1		
Hepato-cholangiocellular carcinoma (H	ICCC)0	0	0	0	0	1	0	2		
Total hepatic tumor	0	0	0	0	0	8(27.5%)	0	10(35.7%)		

Table 4Experiment 2 : summary of findings of liver microscopy.

Criteria for changes in oval cell : ^a mild degree, ^b moderate degree, ^c high degree. Number of animals sacrificed are in the parenthesis.



Fig 4–The liver of a male Wistar control-group rat at 60 weeks.



Fig 5–The liver of a male Wistar DDT-group rat at 60 weeks.

out the liver section. Animals in group 4 (AFB₁/DDT) showed broadly similar changes, however 1 of these 5 rats had developed neoplastic nodules by week 32 (Fig 3).

At the end of the experiment (60 weeks), there were no remarkable changes in the livers of control and DDT-treated rats (Figs 4 and 5). In the AFB_1 -treated group, one rat developed a hepatic tumor mass (Fig 6). Histologically, some livers from rats in this group showed enlargement, vacuolization, fatty change and necrosis of hepatocytes. All rats exhibited massive proliferation of oval cells with large pale nuclei in the periportal zone. Most rats developed altered cell foci. Acidophilic cell foci were

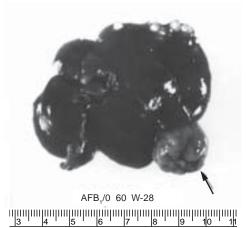


Fig 6–The liver of a male Wistar rat that had been given dietary AFB₁ (4 ppm) for 6 weeks, followed by a basal diet for 12 weeks, and then chow pellets. The arrow points to a tumor of the left central lobe (60 weeks).

frequently seen. Neoplastic nodules and hepatocellular carcinoma and cholangiocellular carcinomas were seen, but in lower numbers and severity compared with the rats that had been exposed to DDT following initiation with AFB₁ (group 4, Fig 7). In the DDT-treated group, some rats showed massive oval cell proliferation in the portal area. Some hepatocytes showed a few distinctive intracytoplasmic inclusion bodies as well as hypertrophy and hyperplasia of Kupffer cells.

Although cholangiocellular carcinomas were rare, one rat in the group that received AFB₁ and one from the AFB₁/DDT group developed both hepatocellular carcinoma (Fig 7A) and cholangiocellular carcinoma (Fig 7B). Hepato-cholangio-cellular carcinomas were found in 1 rat that had received AFB₁ alone, and in 2 that had received AFB₁ following DDT (Fig 7C).

Other pathology

Only a limited study of other tissues was made, since the major objective was to compare liver tumor development. However, the kidneys were examined in view of earlier reports of tumors in these organs in AFB₁-treated rats.

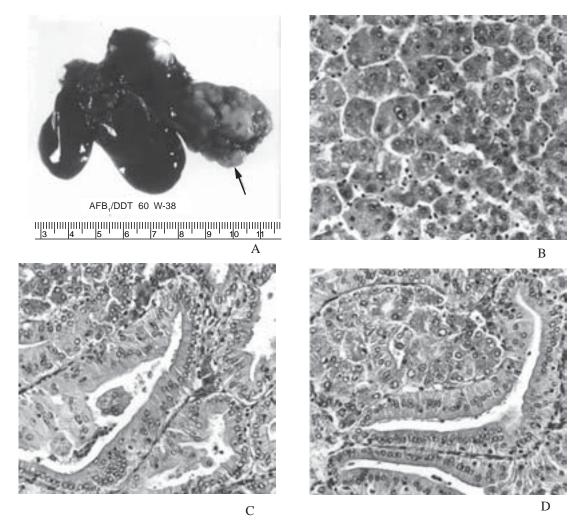


Fig 7–The liver of a male Wistar rat that had been given AFB₁ (4 ppm) for 6 weeks, followed by DDT (500 ppm) for 12 weeks. The arrow points to a large tumor of the left central lobe at 60 weeks (A), showing histologic appearance of hepatocellular carcinoma (B), and cholangio-cellular carcinoma in which neoplastic glands of various sizes and the cells' pappillary projections extend into the lumen (C), and mixed hepatocellular / cholangiocellular carcinomas (D). Hematoxylin and eosin stain, magnification x 600.

The kidneys of rats receiving AFB₁, AFB₁/DDT and of the controls showed varying degrees of nephritis with interstitial fibrosis and chronic inflammation. No renal cell carcinoma was found.

DISCUSSION

In experiment 1, neoplastic nodules were observed in only 1 of the 14 male Buffalo rats

that had been given a single oral dose of AFB_1 (5 mg/kg). This was similar to the result of a previous study in which male Fisher rats were given the same dose of 5 mg/kg toxin (Wogan and Newberne, 1967). In addition, Lemonnier *et al* (1975) found that 9 of 19 female Wistar rats given a single oral dose of 7 mg/kg AFB_1 (the approximate LD_{50}) developed hepatoma at 120 weeks. The present results showed a lower susceptibility to liver cancer as the result of a single oral dose of AFB_1 with DDT than was

observed in an earlier study which was initiated by a brief dose of 2-acetylaminofluorene (2-AAF) (Peraino et al, 1975). Three possible reasons may explain this finding: (1) Buffalo rats are more resistant to hepatocarcinogenesis from AFB, than both Fisher rats (Wogan and Newberne, 1967) and Wistar rats (Angsubhakorn et al, 1990; (2) the subsequent continuous feeding of DDT in our experiment was at a lower dose (100 ppm) than used in other studies (Peraino et al, 1975); (3) the duration of exposure to the DDT was shorter (20 weeks) in this study than in the study of Peraino et al (1975) (41 weeks). Such promoters may have to be given for a period long enough to enable altered cells to develop neoplastic nodules and then hepatocellular carcinoma.

In experiment 2, the percentage incidence of altered foci and neoplastic nodules were not different, but the incidence of carcinoma was higher in group 4 rats (10/28;35.7%), which received 4 ppm AFB, for 6 weeks followed by 500 ppm DDT for 6 weeks, than in group 2 rats (8/29; 27.5%), which were given AFB, alone. The low enhancement of DDT on AFB, observed in this study may have been due to the different route of AFB₁ administration; Rojanapo et al (1988) gave 1.9 mg AFB, orally for 14 weeks followed by 15 weeks of DDT given by intragastric intubation (75 µg, 2 times/ week). Liver tumors were found in 8 of 24 rats given AFB, followed by DDT. In addition, promoter activity was at its maximum when DDT was given one week after the completion of AFB₁ treatment (Rojanapo et al, 1993).

The liver tumor histopathology in our experiment was similar to that reported by Rojanapo *et al* (1988), who found that 2 rats in each of the treatment groups developed mixed hepato-cholangiocellular carcinomas. DDT administration after AFB_1 appeared to promote the formation of carcinomas containing mixed hepato-cholangiocellular elements.

Two mechanisms have been postulated for two-stage hepatocarcinogensis (Williams, 1981): first, a promoter could completely convert the partially transformed cells to fully neoplastic cells, second, the promoter could act on neoplastic cells to enable them to proliferate into an overt neoplasm. The low incidence of liver tumor in this experiment may have been due to an insufficient amount of promoter (Flodstrom *et al*, 1990). Insufficient promotion fails to inhibit intercellular communication and therefore fails to prompt the release of neoplastic cells that grow progressively to form neoplasm. In addition, DDT would have to be administered for a period long enough to enable the altered cells to proliferate into carcinomas.

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REFERENCES

- Alpert ME, Hutt MSR, Wogan GN, Davidson CS. Association between aflatoxin content of food and hepatoma frequency in Uganda. *Cancer* 1971; 28: 253-60.
- Angsubhakorn S, Bhamarapravati N, Romruen K, Sahaphong S. Enhancing effects of dimethynitrosamine on aflatoxin B₁ hepatocarcinogenesis in rats. *Int J Cancer* 1981; 28: 621-6.
- Angsubhakorn S. Effects of aflatoxin on human and animal health in Thailand. *J Thai Vet Med Assoc* 1983; 34: 285-330.
- Angsubhakorn S, Get-Ngern P, Miyamoto M, Bhamarapravati N. A single dose-response of aflatoxin B_1 on rapid liver cancer induction in two strains of rats. *Int J Cancer* 1990; 46, 664-8.
- Cameron GR, Cheng KK. Failure of oral DDT to induce toxic changes in rats. *Br Med J* 1951; 11: 819-21.
- Deichmann WB, Keplinger M, Sala F, Class E. Synergism among oral carcinogen. IV. The simultaneous feeding of four tumorigens to rats. *Toxicol Appl Pharmacol* 1967; 11: 88-103.

- Flodstrom S, Homming H, Warngard L, Ahlborg UG. Promotion of altered hepatic foci development in rat liver, cytochrome P-450 enzyme induction and inhibition of cell-cell communication by DDT and some structurally related organohalogen pesticides. *Carcinogenesis* 1990; 11: 1413-7.
- Fitzhuch OG, Nelson AA. Chronic oral toxicity of DDT. *J Pharmacol Exp Ther* 1947; 89: 18-30.
- Gorchev HG, Jelinek GF. A review of the dietary intakes of chemical contaminants. *Bull WHO* 1985; 63: 945-62.
- Gornall AG, Bardawill CJ, David MM. Determination of serum protein by means of the biuret reaction. *J Biochem* 1948; 77: 751-66.
- Haag HB, Finnegan JK, Larson PS, Reise W, Dreyfuss ML. Comparative chronic toxicity for warmblooded animals of DDT and DMDT (methoxychlor). Arch Int Pharmacodyn 1950; 83: 491-501.
- Kaiser J, Enserink M. Treaty takes a POP at the dirty dozen. *Science* 2000; 290:2053.
- Kitakawa R, Hino O, Nomura K, Sugano H. Doseresponse studies on promoting and carcinogenic effects of phenobarbital and DDT in rat hepatocarcinogenesis. *Carcinogenesis* 1984; 5: 1653-5.
- Klimbrough R, Gaines TB, Sherman JD. Nutritional factors, long-term DDT intake and chloroleukemia in rats. *J Natl Cancer Inst* 1964; 33: 215 -25.
- Klimmer OR. Experimentelle untersuchungen uber toxicologie insecticider chloroerter kohlenwasserstof. Arch Exp Path Pharmakol 1955; 277: 183-95.
- Lemonnier FJ, Scotto JM, Thuong-Trieu V. Influence of riboflavin on disturbances in tryptophan metabolism and hepatoma production after a single dose of aflatoxin B₁. *J Natl Cancer Inst* 1975; 55: 1085-87.
- Montesano R, Hainaut P, Wild CP. Hepatocellular carcinoma: From gene to public health. *J Natl Cancer Inst* 1997; 89: 1844-51.
- Peers FG, Linsell GA. Dietary aflatoxins and liver cancer – a population-based study in Kenya. *Br J Cancer* 1973; 27: 437-84.
- Peers FG, Gilman GA, Linsell CA. Dietary aflatoxins

and human liver cancer. A study in Swaziland. *Int J Cancer* 1976; 17: 167-76.

- Peers FG, Bosch X, Kaldor J, Linsell A, Pluumen M. Aflatoxin exposure, hepatitis B virus infection and liver cancer in Swaziland. *Int J Cancer* 1987; 39: 545-53.
- Peraino C, Fry RJM, Staffeldt E, Christopher JP. Comparative enhancing effects of phenobarbital, amobarbital, diphenylhydantin, and dichlorodiphenyltrichloroethane on 2- acetylaminofluorene-induced hepatic tumorigenesis in the rat. *Cancer Res* 1975; 25: 2884-90.
- Reitman S, Frankel S. A colorimetric method for the determination of serum glutamic oxaloacetic and serum glutamic pyruvic transaminases. *Am J Clin Pathol* 1957; 28: 56-59.
- Rodkey FL. Direct spectrophotometric determination of albumin in human serum. *Clin Chem* 1965; 11: 478-87.
- Rojanapo W, Tepsuwan A, Kupradinum P, Chutimataewin S. Modulation of hepatocarcinogenicity of aflatoxin B₁ by the chlorinated insecticide, DDT. In: Nigam SK, McBrien DCH, Slater TF, eds. Eicosanoids, lipid peroxidation and cancer. Berlin: Springer-Verlag; 1988: 327-38.
- Rojanapo W, Kupradinum P, Tepsuwan A, Tanyakaset M. Effect of varying the onset of exposure to DDT on its modulation of AFB₁-induced hepatocarcinogenesis in the rat. *Carcinogenesis* 1993; 14: 663-7.
- Rossi L, Ravere M, Repetti G, Santi L. Long-term administration of DDT or phenobarbital-Na in Wistar rats. *Int J Cancer* 1977; 19: 179-85.
- Shank RC, Wogan GN, Gibson JB, Nodasuta A. Dietary aflatoxin and human liver cancer II. Aflatoxin in market foods and foodstuffs of Thailand and Hong Kong. *Fd Cosmet Toxicol* 1972a; 10: 61-70.
- Shank RC, Gordon JE, Wogan GN, Nondasuta A, Subhamani B. Dietary aflatoxins and human liver Cancer III. Field survey of rural Thai families for ingested aflatoxins *Fd Cosmet Toxicol* 1972b; 10: 71-84.
- Shank RC, Bhamarapravati N, Gordon JE, Wogan GN Dietary aflatoxins and human liver cancer IV. Incidence of primary liver cancer in two municipal populations of Thailand. *Fd Cosmet Toxicol* 1972c; 10: 171-9.

- Steward HI, William G, Keysser CH, Lombard LS, Montali RJ. Histologic typing of liver tumors in the rat. *J Natl Cancer Inst* 1980; 64: 179-206.
- Squire RA, Levitt MH. Report of a workshop on classification of specific hepatocellular lesions in rats. *Cancer Res* 1975; 35: 3214-23.
- Suwankerd W, Prajakwong S. In : Suwankerd W, Prajakwong S, eds. Study areas for DDT susceptibility in Northern Thailand. Chiang Mai: So Sap Karnpim, 1995.
- Treon JR, Cleveland FD. Toxicity of certain chlorinated hydrocarbon insecticides for laboratory animals, with special reference to aldrin and dieldrin. *J Agri Fd Chem* 1955; 3: 402-5.
- Van Rensburg SJ, Cook Mozaffari P, van Schalkwyk

DJ, *et al*. Hepatocellular carcinoma and dietary aflatoxin in Mozambique and Transkei. *Br J Cancer* 1985; 5 : 713-26.

- Wissermann M, Trishnananda L, Tomatis L, *et al*, Storage of organochlorine insecticides in the adipose tissue of people from Thailand. *Southeast Asian J Trop Med Public Health* 1972; 3: 280-5.
- Williams GM. Liver carcinogenesis : The role for some chemicals of an epigenetic mechanism of liver tumor promotion involving modification of the cell membrane. *Fd Cosmet Toxicol* 1981; 19: 577-83.
- Wogan GN, Newberne PM. Dose-response characteristic of aflatoxin B₁ carcinogenesis in rat. *Cancer Res* 1967; 27: 2370-6.