RAPID DIAGNOSIS OF HELICOBACTER PYLORI INFECTION IN GASTRIC IMPRINT SMEARS

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Abstract. The objective of this study was to determine the sensitivity, specificity, positive (PPV), and negative predictive values (NPV) of Diff-Quik-stained gastric imprint cytology smears in the detection of H. pylori compared with histology. Air-dried imprint smears of gastric biopsies from 150 patients were stained by the Diff-Quik method in the endoscopy suite and examined for H. pylori, providing results within minutes. The presence of inflammation and intestinal metaplasia were documented. The same biopsy was processed and stained with H&E and Warthin-Starry stains, and reviewed by a different pathologist blind to the imprint cytology results. Ninety-four of the 150 patients were male with a mean age of 50 years. Based on histology, the H. pylori prevalence was very low at 8%. The sensitivity and specificity of imprint cytology in the detection of H. pylori were 83.3% and 100%, respectively. The PPV and NPV were 100% and 98.6%, respectively. There were two false negatives and no false positives. A combination of imprint cytology and histology achieved 100% sensitivity. Imprint smears did not provide added value over histology with regards to inflammation and metaplasia. Gastric imprint smears stained with Diff-Quik method is a rapid, cheap, and reliable method for the detection of H. pylori and have their best results when complemented with histology.

INTRODUCTION

Helicobacter pylori colonization of the gastric mucosa is associated with the pathogenesis of gastritis, peptic ulcer disease, and gastric malignancy. As eradication of this organism has become part of clinical practice, much research has been done in assessing the sensitivity and reliability of the available diagnostic methods for the detection of this organism (Loffeld et al., 1993). Though culture is the gold standard, it lacks sensitivity, is technically difficult and costly. Urease tests and histological examination of gastric specimens are frequently used methods in this country, while histology is the only method employed in our institution with a turnaround time of several days.

Cytology of gastric brushings and imprint smears have been described as reliable methods for the detection of H. pylori (Mason et al., 1989; Pinto et al, 1991; Debrongnie et al., 1992; Mendoza et al., 1993; Carmona et al., 1995; Rodriguez et al., 1995; Faraker, 1996; Rey et al., 1997; Trevisani et al 1997; Cubukcu et al., 2000). The Papanicolaou and May-Grunwald-Giemsa staining methods are widely used and the procedures take about 30 minutes. The Diff-Quik staining method is even more rapid, cheap, and can be performed in the endoscopy suite with the identification of H. pylori within minutes.

The aim of this study was to determine the sensitivity, specificity, positive (PPV) and negative predictive values (NPV) of Diff-Quik-stained gastric imprint smears in the detection of H. pylori compared with histology of the same biopsy used for imprint. It was also hoped that its usefulness, as a diagnostic method, could be determined in this region of the country, an area with a low prevalence of H. pylori infection (Uyub et al., 1994; Kaur and Raj, 2002). The value and agreement of signs of inflammation and intestinal metaplasia on the imprint, compared with histology, were also of interest.
MATERIALS AND METHODS

A total of 150 patients undergoing esophago-gastro-duodenoscopy (OGDS) for upper gastrointestinal symptoms were included in the study, which had prior approval from the ethics committee of this institution. Exclusion criteria included patients undergoing OGDS as an emergency procedure and in which stomach biopsies were not indicated such as esophageal and other systemic diseases. During endoscopy, three gastric biopsies are usually taken, two from the antrum and one from the body. In this study, only one of the gastric biopsy specimens, preferably from the antrum, was rolled gently on a clean glass slide using forceps to make an imprint smear, before placing it in 10% formalin for subsequent histopathological examination. Only one gastric imprint smear per patient was prepared. The smears were air-dried and stained by the Diff-Quik (Lab Aids Pty, Malaysia) method by a pathologist in the endoscopy suite. This staining method took an average of 2 minutes. The smears were then air-dried, mounted and examined for *H. pylori* under 400x magnification. The bacteria were identified by their spiral or S shaped morphology and the purple-violet color within the well-preserved gastric mucus. The presence of neutrophils, large numbers of lymphocytes, lymphoid aggregates and goblet cells reflecting intestinal metaplasia were documented when present.

The gastric biopsy was processed in the usual way for histology and stained with hematoxylin and eosin (H&E) and Warthin-Starry stains for *H. pylori* detection. The slides were reviewed by a different pathologist, blinded to the imprint cytology results. Besides identification of *H. pylori*, other parameters, such as grade of chronic inflammation, activity, and intestinal metaplasia, were also determined.

The results of the imprint smears in the detection of *H. pylori* were compared to histology. The sensitivity, specificity, negative and positive predictive values were calculated. As for the other variables, such as inflammation and metaplasia, imprint smears and histology were compared.

RESULTS

Ninety-four of the 150 patients were male with a mean age of 50 years (range 11 to 84 years). The ethnic composition of the patients was as follows: Malay 110 (73.3%), Chinese 28 (18.7%), Indian 6 (4%), and ‘other races’ 6 (4%) which comprised 2 Thai, 2 Pakistani, 1 Burmese and 1 Iraqi.

The Diff-Quik staining method was simple and rapid, taking about 2 minutes. Imprint smears produced good cytologic preparations. The cellular content was adequate and gastric mucus stained a light violet. Only two smears had poor cellularity. *H. pylori* were easily visualized within the gastric mucus (Fig 1). Although other bacteria were also stained by the Diff-Quik method, the characteristic spiral shape of *H. pylori* proved its positive identification.

The gastric biopsy specimens from which

Fig 1–*H. pylori* and gastric epithelial cells in gastric imprint cytology (Diff-Quik, x 400).
imprint smears were made retained good quality and the histological examination was not compromised in any way.

Based on histology, the overall *H. pylori* prevalence in this endoscoped population was 8% (12/150). The frequency of *H. pylori* infection among the endoscoped population according to the ethnic distribution was as follows: Malay 3/110 (2.7%), Chinese 7/28 (25%), Indian 1/6 (16.7%), and one Burmese.

The sensitivity and specificity of imprint cytology compared to histology for the identification of *H. pylori* were 83.3% and 100%, respectively. PPV was 100% and NPV was 98.6% (Table 1). There were two false negative cases. Two positive cases on imprint smear, that were initially diagnosed as negative on histology, showed a low density of *H. pylori* on review. The sensitivity, specificity, PPV and NPV of 100% were achieved if imprint cytology and histology were combined.

There was very poor agreement between imprint smears and histology with regards to inflammation, and intestinal metaplasia. The presence of lymphocytes or lymphoid aggregates on imprint smears did not correspond to chronic inflammation or its severity on histology. Goblet cells were not identified on imprint smears and noted on only 2 biopsies in the *H. pylori*-positive cases. Of the 12 subjects with *H. pylori* infection, only 2 showed the presence of neutrophils on imprint smears while activity was seen in 11 cases on histology.

**DISCUSSION**

The available methods for the detection of *H. pylori* organisms are many and broadly categorized as invasive and non-invasive methods (Mason *et al*, 1989; Pinto, *et al* 1991; Debongnie *et al*, 1992; Loffeld *et al*, 1993; Mendoza *et al*, 1993; Carmona *et al*, 1995; Rodriguez *et al*, 1995; Faraker, 1996; Rey *et al*, 1997; Trevisani *et al*, 1997; Cubukçu *et al*, 2000). Invasive methods that require gastrointestinal endoscopy and biopsy of the stomach include culture, histology, urease tests, and cytology. Non-invasive techniques comprise breath tests and serology. Histology is often accepted as the ‘gold standard’ in diagnosing *H. pylori* infection as culture has marked limitations. Information with regards to histopathology of the gastric mucosa is an important advantage. As *H. pylori* is found in the gastric mucus on the surface epithelium and gastric foveolae, cytology of imprint smears and brushings have proven to be reliable methods for its detection (Pinto *et al*, 1991; Debongnie *et al*, 1992; Mendoza *et al*, 1993; Carmona *et al*, 1995; Rodriguez *et al*, 1995; Faraker, 1996; Rey *et al*, 1997; Trevisani *et al* 1997; Cubukçu *et al*, 2000). Previously reported studies used various combinations of techniques and staining methods. Brushings and touch imprints were common, while the stains used included Papanicolaou, May-Grunwald-Giemsa, Hema-Gurr and Diff-Quik. We decided that performing imprint smears of gastric biopsy specimens before routine histological processing added no extra procedure or inconvenience to the endoscopist or patient. Comparing imprint smears and matching histology was ideal in eliminating sample bias.

Imprint smears provided good cellularity and one smear per patient was adequate. Epithelial and inflammatory cells, as well as *H. pylori*, were easily visualized with the Diff-Quik stain. Other bacteria often present in de-acidified stomachs may be confused with *H. pylori* stain. Other bacteria often present in de-acidified stomachs may be confused with *H. pylori*. However, the characteristic morphology of *H. pylori* can be readily made at 400x magnification.

The Diff-Quik staining procedure is simple and rapid, requiring no additional staff besides the pathologist who stained and interpreted the imprint smears. The turnaround time was an average of 10 minutes, compared with three to five days for a histological report. This provides a tremendous advantage, as therapy can be commenced before the patient leaves the endoscopy suite on the same day.
The sensitivity of *H. pylori* detection in brush or imprint cytology using various stains has been reported to be between 71% and 100%, while the specificity was 90% to 100% (Pinto *et al* 1991; Mendoza *et al*., 1993; Carmona *et al*., 1995; Faraker, 1996; Rey *et al*., 1997; Trevisani *et al*., 1997; Cubukcu *et al*., 2000). In our study, the sensitivity and specificity of imprint cytology compared with histology were 83.3% and 100%, respectively. The PPV was 100% and NPV was 98.6%, which is comparable to another study quoting the PPV as 93.8% and NPV as 94.6% using Diff-Quik-stained gastric antral brushings (Carmona *et al*., 1995). False negatives can arise when the bacterial load is low, as was seen in two of our subjects. A review of the imprint smears confirmed they were negative, while the biopsies revealed low density and patchy distribution of the organism. On the other hand, two *H. pylori*-infected patients were identified by imprint smears, but not by initial histology (H&E and Warthin-Starry). Careful scrutiny of the biopsies showed a low and patchy bacterial load. The density of *H. pylori* organisms seen on imprint smears did not reflect that seen in biopsies. In subjects with a low bacterial load, imprint smears and gastric brushings probably provide better sampling of *H. pylori*, which reside in the gastric surface mucus layer. The brushing technique has the disadvantage of requiring an added procedure. Variable loss of the mucus layer is unavoidable during histological processing. The sensitivity in our study is affected by the low bacterial load and, to a certain extent, the low prevalence of *H. pylori* in the study population. However, imprint cytology, combined with histology, achieved 100% sensitivity.

Though the Warthin-Starry stain was performed to facilitate identification of *H. pylori*, it is expensive and in the hands of experienced pathologists, can be omitted.

Inflammatory changes on imprint smears did not correlate well with histology and, therefore, did not provide any added value, though it was considered useful in a previous study (Pinto *et al*, 1991). It is clear that histological examination is necessary to provide information on mucosal inflammation, metaplasia, atrophy and other histopathological changes.

The quality of the gastric biopsies from which the imprints were made was not adversely affected, and proper histological examination could be done, validating previous studies (Debongnie *et al*, 1992; Cubukcu *et al*., 2000). With this advantage, imprint cytology can easily complement histological examination with minimal added cost.

The *H. pylori* infection prevalence rate of 8% among an endoscoped population is unusually low but it is in concordance with previous studies done in this region of the country (Uyub *et al*, 1994; Kaur and Raj, 2002). The interesting ethnic distribution of *H. pylori* infection, being very low in Malays (2.7%) compared with non-Malays (20%), is a commonly reported phenomenon in this multiracial country (Goh, 1997; Goh and Parasakhti, 2001). The predominant Malay population in this part of the country lowers the prevalence rates even further. This being so, the value and cost effectiveness of imprint cytology is questionable, when taking into consideration the whole patient population undergoing endoscopy. However, with the higher index of suspicion of *H. pylori* infection among the non-Malays, imprint cytology may prove to be of great added value as infected patients can begin therapy immediately.

In conclusion, air-dried gastric imprint smears stained by the Diff-Quik method provide a simple, rapid, cheap, and reliable method for the detection of *H. pylori*, with the enormous advantage of immediate commencement of eradication therapy. This technique can achieve maximum sensitivity with added useful information when combined with histological examination, especially in ethnic groups that have a higher prevalence of the infection.

ACKNOWLEDGEMENTS

This work was supported by the Universiti Sains Malaysia short-term research grant, 304/PPSP/6131180. The authors have no connection to any companies or products mentioned in this article.

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