# SEROLOGIC DETECTION OF *TOXOPLASMA GONDII* INFECTION IN RATTUS SPP COLLECTED FROM THREE DIFFERENT SITES IN DASMARIÑAS, CAVITE, PHILIPPINES

Cristina C Salibay<sup>1</sup> and Florencia G Claveria<sup>2</sup>

<sup>1</sup>Biological Sciences Department, College of Science, De La Salle University-Dasmariñas, Dasmariñas, Cavite; <sup>2</sup>Biology Department, College of Science, De La Salle University-Manila, Taft Avenue, Manila, Philippines

**Abstract**. Acute and chronic cases of toxoplasmosis in *Rattus norvegicus* and *Rattus rattus mindanensis* caught in agricultural, commercial and residential sites in Dasmariñas, Cavite, Philippines were determined serologically. Fifty-eight percent of *R. norvegicus* and 42.0% of *R. r. mindanensis* were positive for anti-*T. gondii* antibodies (Abs). Infection was higher in male rats, and those caught in the commercial site had 100.0% seropositivity. Thirty percent of the *R. norvegicus* and 51.0% *R. rattus mindanensis* had acute infection, with 1:64-1:128 Abs titer. Seventy percent of the *R. norvegicus* and 49.0% of *R. rattus mindanensis* were chronically-infected with Abs titer 1:256-1:2048 and 1:256-1024, respectively. The association between the presence of infection with the rat gender and species and their collection sites was insignificant (p>0.05). In a related study, however, mice experimentally-inoculated brain tissue homogenate obtained from chronically-infected *Rattus* spp, manifested differences in the onset as well as, severity of infection which was histopathologically evaluated, suggestive of a possible difference in *T. gondii* parasite strain(s) infecting different rat populations.

## INTRODUCTION

Rattus species are the most diverse among the rodents, of which 20 species are considered important pests (Fall, 1980). In the Philippines, the two most important pestiferous rats that survive and proliferate around human habitation include Rattus norvegicus and Rattus rattus mindanensis (Fernando et al, 1985). Besides the agricultural and domestic damages caused by rats, they are also carriers of human disease, including toxoplasmosis (Morse, 1956; Tenter et al, 2000). Toxoplasma gondii, a zoonotic, heteroxenous obligate intracellular parasite has developed several potential routes of transmission within and between different host species (Levine, 1973; Ferguson et al, 1999). Common species of domestic and urban rats are chronic carriers of Toxoplasma and serve as a potential reservoir of infection to cats as well as other livestock animals (Wallace, 1973; Webster and MacDonald, 1995; Battersby, 1998). In the absence of cats, T. gondii can be maintained by vertical transmission in rats (Dubey, 1997a; Webster et al, 1994). Nevertheless, Wastling et al (2000) underscored the possibility of the natural life cycle of T. gondii via a cat-to-rodent-to-cat transmission which may indiscriminately involve infection of other warm blooded animals.

Correspondence: Cristina C Salibay, Biological Sciences Department, College of Science, De La Salle University-Dasmariñas, Dasmariñas, Cavite, Philippines.

E-mail: ccsalibay@mail.dasma.dlsu.edu.ph

In the Philippines, documented studies on toxoplasmosis are largely serologic in nature, in swine (Mendoza, 1974; Manuel and Tubongbanua, 1977; Marbella, 1980; Manuel, 1982), cats (Minervini, 1985; Dans, 2002), and in a few selected communities in Metro Manila, Mindoro and Leyte (Kawashima et al, 2000). Taking into account the wide distribution and abundance of rodents in environments close to human habitation (Fernando et al, 1985; Gratz, 1988), and their role as carriers/reservoir hosts of T. gondii (Galuzo, 1970; Fall, 1980), the present study sought to establish serologically the presence of T. gondii infection in R. norvegicus and R. rattus mindanensis caught in agricultural (AGR), commercial (COM), and residential (RES) sites in Dasmarinas, Cavite, Philippines.

## MATERIALS AND METHODS

Using spring-door wire traps with food as bait, *Rattus* species were collected from AGR, COM and RES areas of the municipality of Dasmariñas, Cavite. *Rattus norvegicus* and *R. rattus mindanensis* were then identified (Fernando *et al*, 1985). Dasmariñas, Cavite is situated in the southern part of the Island of Luzon, Philippines, and is approximately 57 kms from Metro Manila, the National Capital Region (Research, Statistics, Monitoring and Evaluation Division, 2001). In the past ten years, the construction of commercial establishments and subdivisions has resulted in massive industrialization of the area and necessitated the conversion of a wide area of agricultural land. Five ml of blood sample was extracted from each rat through venipuncture of the jugular vein. Blood was allowed to clot at RT for 30 minutes and centrifuged at 1,500 rpm for one minute. Sera were transferred into properly labeled tubes, stored in a refrigerator ( $4^\circ$ - $8^\circ$ C), and were serologically processed within 24 hours post-collection.

Rat sera were assayed for the presence of anti-*T.* gondii Abs using the TOXOCELL AD Direct Agglutination Test Kit. The test kit contained a suspension of highly purified and concentrated *Toxoplasma* (Ags) used to determine the presence of IgM Abs (1:64-1:128) indicative of acute infection, and IgG Abs ( $\geq$ 1:256) indicative of chronic infection.

Serologic data generated per rat species and collection site were statistically analyzed using chisquare analysis and one-way analysis of variance (ANOVA) ( $p \le 0.05$ ).

#### RESULTS

A total of 157 rat sera were assayed for anti-*T.* gondii Abs. Eighty-seven (55.0%) were seropositive (sero<sup>+</sup>), broken down as follows: 50 (60.0%) of the 83 *R. norvegicus*, and 37 (50.0%) of the 74 *R. rattus* mindanensis (Table 1). While serologic data suggest greater susceptibility of *R. norvegicus* to *T. gondii* relative to *R. rattus* mindanensis, statistical analysis showed insignificant association between parasite infectivity, rat species, and collection sites.

More male rats tested sero<sup>+</sup>, except for *R. rattus* mindanensis caught in the COM site (Fig 1). Statistical analysis of gender-related data on toxoplasmosis in *R.* norvegicus and *R. rattus mindanensis* revealed insignificant association (p>0.05).

Comparison of acute and chronic cases across two

rat species and three collection sites is summarized in Fig 2. Thirty-five (70.0%) of the sero<sup>+</sup> *R. norvegicus*, and 18 (49.0%) of sero<sup>+</sup> *R. rattus mindanensis* were chronic cases and registered anti-*T. gondii* Abs titer of 1:256-1:2048, and 1:256 1:1024, respectively (Table 2). All sero<sup>+</sup> COM-site *R. norvegicus* were chronically-infected.

### DISCUSSION

The relatively high (>55.0%) number of sero<sup>+</sup> rats is consistent with earlier documented studies in domestic and wild rats (Webster and MacDonald, 1995; Battersby, 1998). Although more male rats tested sero<sup>+</sup> but there was insignificant association (p>0.05) of gender-related toxoplasmosis and this finding corroborates earlier findings in cats (Minervini, 1985) and humans (Lee *et al*, 2000), suggestive of the parasite's indiscriminate infectivity to both genders (Minervini, 1985; Dans, 2000).

Present findings are consistent with earlier reports that have identified domestic rats as chronic carriers of the tissue form of *Toxoplasma* (Wallace, 1973; Sasaki *et al*, 1976; Dubey *et al*, 1997b) and as a potent reservoir of infection to cats (Wallace, 1973).

The wide range of anti-*T. gondii* IgG titers assayed in the present study is suggestive of a difference in the status/persistence of *T. gondii* infection in rats surveyed vis-à-vis the frequency of their re-exposure to infection. Galuzo (1970) pointed out that the potential of animals to become reservoir hosts of *T. gondii* increases with re-exposure, where the host immune state is heightened with an increase in anti-*T. gondii* IgG titer. However, considering the continued proliferation of parasites even in the presence of high Abs titer, humoral-related immunity may still be insufficient to provide host protection (Stites *et al*, 1984).

Table 1

Number and serology of *R. norvegicus* and *R. rattus mindanensis* collected from agricultural (AGR), commercial (COM), and residential (RES) sites.

Collection site	No. sero+ <i>R. norvegicus/</i> total no. assayed	No. sero+ <i>R. rattus</i> <i>mindanensis/</i> total no. assayed (%))	Total no. sera (%)	
AGR	20/34	13/28	33/62 (53.0)	
СОМ	16/24	8/16	24/40 (60.0)	
RES	14/25	16/30	30/55 (54.0)	
Total sero+/total no. of sera s	pecies (%) 50/83 (60.0)	37/74 (50.0)	87/157 (55.0)	

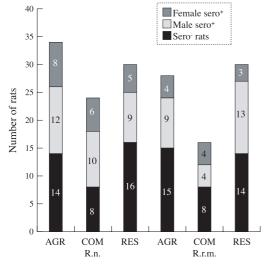


Fig 1- Gender-related *T. gondii* seropositivity differences in *R. norvegicus* (R.n.) and *R. rattus mindanensis* (R.r.m.) caught in agricultural (AGR), commercial (COM), and residential (RES) sites.

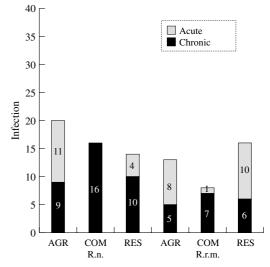


Fig 2- Comparison of acute and chronic cases of *T. gondii* infection in *R. norvegicus* and *R. rattus mindanensis* caught in AGR, COM, and RES sites; antibody titer (acute: 1:64-1:128; chronic: 1:256-1:2048).

Table 2							
Toxoplasma gondii chronically-infected R. norvegicus and R. rattus mindanensis and							
titers of IgG antibodies.							

Species	Collection site	Titer (IgG Abs)			Total chronic cases(%)		
		1:256	1:512	1:1024	1:2048	Per site	Per species
R. norvegicus	AGR	3	4	2	0	9	35/50 (70.0)
	COM	5	7	3	1	16	
	RES	5	2	1	2	10	
R. rattus	AGR	2	2	1	0	5	18/37 (49.0)
mindanensis	COM	4	1	2	0	7	
	RES	1	3	2	0	6	
Total no. of rats per IgG titer (%)		20	19	11	3	53/87 (61.0)	

In conclusion, we have established serologically the presence of *T. gondii* infection in *R. norvegicus* and *R. rattus mindanensis* caught in the AGR, COM, and RES sites in Dasmarinas, Cavite, Philippines. To our knowledge, the present findings may represent the first documented study of *T. gondii* infection in rats in the country.

#### REFERENCES

Battersby S. Urban rat control, underground drainage and public health: report of a post survey pest control companies in England and Wales. Guildford, UK: University of Surrey, 1998:2-3.

Dans BK. A study on the presence of *Toxoplasma* gondii antibodies in the blood serum of stray cats in the Ninoy Aquino Parks and Wildlife Nature Center using the immunocomb enzyme-linked immunosorbent assay feline *Toxoplasma* and *Chlamydia* antibody test kits. Los Baños, Laguna: University of the Philippines, Los Baños (UPLB), 2002. Thesis.

Dubey JP. Toxoplasmosis in rats (Rattus norvegicus):

congenital transmission to first and second generation offsprings and isolation of *Toxoplasma gondii* from seronegative rats. *Parasitology* 1997a;115:9-14.

- Dubey JP. Distribution of tissue cysts in tissues of ratfed oocysts. *J Parasitol* 1997b; 83:755-7.
- Fall MW. Management strategies for rodent damage problems in agriculture. In: Proceedings of the Symposium on Small Mammals: Problems and Control. *BIOTROP Special Publication* 1980;12: 177-82.
- Ferguson DJP, Birch-Andersen A, Siim JC, Hutchison WM. Observations on the ultrastructure of the sporocyst and the initiation of sporozoite formation in *Toxoplasma gondii*. Acta Path Micro Scand B 1999;87:183-90.
- Fernando FS, Benigno EA, Hoque MM, et al. Rodent biology and control (with special reference to the Philippines). Los Banos, Laguna: University of the Philippines, 1985:152 pp.
- Galuzo IG. The epizootiology of toxoplasmosis. In: Toxoplasmosis of animals. 1<sup>st</sup> ed. College of Veterinary Medicine, University of Illinois, 1970: 395-413.
- Gratz NG. Rodents and human disease: a global appreciation. In: Rodent pest management. Boca Raton: CRC Press, 1988:45-9.
- Kawashima T, Khin-Sane-Win T, Kawabata M, Barzaga N, Matsuda H, Honishi E. Prevalence of antibodies to *Toxoplasma gondii* among urban and rural residents in the Philippines. *Southeast Asian J Trop Med Public Health* 2000;31:742-6.
- Lee Y-H, Noh H-J, Hwang O-S, Lee S-K, Shin D-W. Seroepidemiological study of *Toxoplasma gondii* infection in the rural area Okcheon-gun, Korea. *Korean J Parasitol* 2000;38:251-6.
- Levine ND. Protozoan parasites of domestic animals and of man. 2<sup>nd</sup> ed. Minnesota: Burgess Publication, 1973:245 pp.
- Manuel MF. Prevalence of *Toxoplasma gondii* antibodies in swine in the Philippines. *Phil J Vet Med* 1982;20:71-7.
- Manuel MF, Tubongbanua R. A serological survey on the incidence of toxoplasmosis among slaughtered

pigs in Metro Manila. *Phil J Vet Med* 1977;16:9-19.

- Marbella CO. *Toxoplasma gondii*: antibody titer determination, histopathologic and isolation studies from naturally infected swine. Los Banos, Laguna; University of the Philippines, Los Banos (UPLB), 1980. Thesis.
- Mendoza CB. A serological survey on the incidence of toxoplasmosis in swine. Los Banos, Laguna: University of the Philippines, Los Banos (UPLB), 1974. Thesis.
- Minervini NA. Study on the presence of *Toxoplasma* gondii antibodies in Philippine-raised cats using the enzyme linked immunosorbent assay test. Los Banos, Laguna: University of the Philippines, Los Banos (UPLB), 1985. Thesis.
- Morse J. Some protozoan diseases of man and animals: anaplasmosis, babesiosis and toxoplasmosis. *Ann NY Acad Sci* 1956:155-233.
- Research, Statistics, Monitoring and Evaluation Division, Provincial Planning and Development Office Trece Martirez, Philippines, 2001.
- Sasaki S, Iida T, Tsuchiya Y, et al. A collective outbreak of toxoplasmosis in swine. J Jpn Vet Med Assoc 1976;29:77-82.
- Stites DP, Stobo JD, Fudenberg HH, Wells VV. Basic and clinical immunology. California: Lange Medical Publications, 1984:99-100.
- Tenter AM, Heckeroth AR, Weiss LM. Toxoplasma gondii: from animals to humans. Aus Soc Parasit 2000;3:34-46.
- Webster JP, Brunton CFA, MacDonald DW. Effects of *Toxoplasma gondii* upon neophobic behavior in wild brown rats, *Rattus norvegicus*. *Parasitology* 1994;109:37-43.
- Webster JP, MacDonald DW. Parasites of wild brown rats (*Rattus norvegicus*) on UK farms. *Parasitology* 1995;111:247-53.
- Wallace GD. The role of the cat in the natural history of *Toxoplasma gondii*. Am J Trop Med Hyg 1973;22:313-22.
- Wastling J, Heap S, Ferguson D. Toxoplasma gondii keeping our guests under control. Biologist 2000;47:234-8.