NEW FORM OF *PARAGONIMUS WESTERMANI* DISCOVERED IN THAILAND: MORPHOLOGICAL CHARACTERISTICS AND HOST SUSCEPTIBILITY

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Abstract. During an intensive field survey for *P. westermani* in southern Thailand, a new form of *Paragonimus* metacercariae was isolated. In this study, we referred to this new form as *P. westermani*-like, as it was almost identical to *P. westermani* in shape. To investigate the susceptibility of feline host to *P. westermani*-like, as well as its morphology at the adult stage, we inoculated the peritoneal cavity of a cat with 60 *P. westermani*-like metacercariae. Morphological examination revealed that the adult *P. westermani*-like recovered from the lungs had a six-lobed ovary, a spermatozoa-filled seminal receptacle, and singly spaced cuticular spines. These findings indicated that the morphological features of *P. westermani*-like were fundamentally identical to those of *P. westermani* (diploid type) at the adult stage. The susceptibility of feline hosts to *P. westermani*-like, we have been investigating the phylogenetic relationships between *P. westermani*-like and *P. westermani* in southern Thailand.

INTRODUCTION

Paragonimus westermani is widely distributed in Asia (Miyazaki, 1991). Individuals from different geographical regions (or countries) show variations in animal and/or human susceptibility, although they share almost identical morphological features at both the adult and metacercarial stages (Blair *et al*, 1998). This implies that they form a complex of cryptic species (Blair *et al*, 1997).

During an intensive field survey for *Paragonimus* in southern Thailand (Rangsiruji *et al*, 2005), we collected another form of *Paragonimus westermani* metacercariae from freshwater crabs, *Phricotelphusa aedes*. These crabs simultaneously acted as the second intermediate host of *P. westermani*. Metacercariae of newly isolated *Paragonimus* were almost identical to those of *P. westermani* in shape, but were much smaller. For descriptive purposes, we refer to this new form as *P. westermani*-like.

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Tel: +81-3-5285-1111; Fax: +81-3-5285-1173 E-mail: hsugi@nih.go.jp In this study, we inoculated a cat with *P. westermani*-like metacercariae isolated from freshwater crabs, *Phricotelphusa aedes*, in order to identify the susceptibility of feline hosts. The morphological characteristics of *P. westermani*-like at the adult stage were also compared with those of *P. westermani*.

MATERIALS AND METHODS

Identification of freshwater crabs

The captured freshwater crabs, belonging to the family Potamidae, were identified as *Phricotelphusa aedes* according to the method of Naiyanetr (1988).

Isolation of Paragonimus metacercariae

Between January and May 2003, we collected 922 freshwater crabs, *Phricotelphusa aedes*, from mountain streams in the Phanom District of Surat Thani Province, Thailand. We examined the crabs for metacercariae, as described previously (Rangsiruji *et al*, 2005). Isolated metacercariae were placed on glass slides and gently pressed under a coverglass for morphological observation and measurement.

Worm recovery from test animal

We inoculated the peritoneal cavity of a cat

with 60 *P. westermani*-like metacercariae. The cat was then treated with prednisolone (20 mg/kg) at 7-day intervals and was necropsied 148 days after inoculation. We examined the whole body of the cat for worms, as described previously (Sugiyama *et al*, 1984). Recovered worms were pressed between two glass slides, fixed in 70% ethanol, stained with borax carmine, and mounted with Canada balsam for morphological observation and measurement.

DNA amplification and sequencing of ITS2 region

We prepared DNA samples from individual *P. westermani* and *P. westermani*-like metacercariae (five metacercariae each). The ITS2 region of the nuclear ribosomal DNA was amplified by PCR and sequenced, as described previously (Sugiyama *et al*, 2002). The primers used were 3S: 5'-GGTACCGGTGGATCACTCGGCTCGTG-3' (forward: Bowels *et al*, 1995) and A28: 5'-GGGATCCTGGTTAGTTTCTTTTCCTCCGC-3' (reverse: Blair *et al*, 1997). We aligned and compared sequences using GENETYX-WIN software (ver 7.0, Software Development, Tokyo, Japan).

RESULTS

New crab intermediate host of *Paragonimus* in southern Thailand

We captured 922 freshwater crabs (Fig 1) from mountain streams in the Phanom District of Surat Thani Province. The crabs were positive for *P. westermani* metacercariae; this is the first report of this crab species serving as a second intermediate host of *P. westermani*. *P. westermani*-like metacercariae were also isolated from the same crab species captured at the same sites.

Morphology of *P. westermani*-like metacercariae from crabs

We isolated 89 *P. westermani*-like metacercariae from the crabs. All were spherical in shape and had thin walls (Fig 2). The thickness of the cyst wall in 30 specimens ranged from 4-14 μ m, with an average of 8.7 μ m. The longitudinal and transverse diameters of the cyst ranged from



Fig 1- Freshwater crabs, *Phricotelphusa aedes*, which serve as the second intermediate host of both *P. westermani* and *P. westermani*-like in southern Thailand.



Fig 2- Photomicrograph of fresh *P. westermani* metacercaria. Bar is 100 μm.



Fig 3- Photomicrograph of fresh *P. westermani*-like metacercariae. Bar is 100 µm.



Fig 4- Adult worm of *P. westermani*-like from a cat inoculated with metacercariae. The worm had six-lobed ovary and singly spaced cuticular spines.

Pw1		060
Pw2	TGTCGATGAAGAGCGCCAGCCAACTGTGTGTGAATTAATGCGAACTGCATACTGCTTTGAACA	060
PwL		060
Pw1	ATATTGCGGCCACGGGTTAGCCTGTGGCCACGCCTGTCCGAGGG	120
Pw2	TCGACATCTTGAACGC	120
PwL		120
Pw1	TCGGCTTATAAACCATCGCGACGCCCAAAAAGTCGCGGCTTGGGTTTTGCCAGCTGGCGT	180
Pw2		180
PwL	······G···T···························	180
Pw1	GATCTCCCCAATCTGGTCTTGTGCCTGTGGGGTGCCAGATCTATGGCGTTTCCCTAACAT	240
Pw2		240
PwL		240
Pw1	ACTCGCGCGCACCCACGTTGCGGCTGAAAGCCTTGACGGGGATGTGGCAACGGAATCGTG	300
Pw2		300
PwL	·····G······	300
Pw1	GCTCAGTAAATGATTTATGTGCGCGTTTCGCTGTCCTGTCTTCATCTGTGGTTCATGTTG	360
Pw2		360
PwL	G.TGT	360
Pw1	CGCGTGGTCTGCGTTCGATGCTGACCTACGTATGTGCCATGTGGTCCATTCTTCTGACCT	420
Pw2		420
PwL		420
Pw1	CGGATTAGACGTGAGTACC 463	
Pw2		
PwL	C	

Fig 5- Aligned sequences of the ITS2 region from *P. westermani* strain Thailand (AF159604, Pw1), *P. westermani* (Pw2) and *P. westermani*-like (PwL). Identical bases are represented by dots. Hyphen indicates missing data. Numbers refer to nucleotide sequence length.

212-252 μ m and from 204-240 μ m, respectively, with an average of 227 x 221 μ m.

The metacercariae of *P. westermani*, also isolated from the same crab hosts, were spherical in shape and had thick walls (Fig 3). The thickness of the cyst wall in five specimens ranged from 19-37 μ m, with an average of 28.2 μ m. The longitudinal and transverse diameters of the cysts ranged from 458-510 μ m and from 438-501 μ m, respectively, with an average of 492 x 480 μ m.

Morphology of an adult worm

On postmortem examination of the test cat, 148 days after inoculation, 13 worms were recovered; 2 from the lungs (being paired in the worm cyst), 2 from the pleural cavity, and 9 from the liver. The worms from the lungs and pleural cavity were identified as either adults (one each, with eggs in the uterus) or pre-adults (without eggs), while the worms from the liver remained in the juvenile stage.

The size of the adult worm from the lung was 3.95 mm in length and 2.83 mm in width. The transverse diameters of the oral and ventral suckers measured 504 µm and 500 µm, respectively. The adult worm had a six-lobed ovary and singly spaced cuticular spines (Fig 4). The seminal receptacle was filled with spermatozoa.

ITS2 sequence analysis

The ITS2 region was amplified from DNA samples of individual P. westermani and P. westermani-like metacercariae using the consensus primers 3S and A28. Sequence analysis of the PCR products revealed that the aligned ITS2 region was 463 bp in length for both P. westermani and P. westermani-like samples. Pairwise comparison of the sequences showed 13 (2.8%) nucleotide differences (Fig 5). Similarity searches of the nucleotide databases GenBank/ EMBL/DDBJ revealed that the ITS2 sequences of P. westermani were identical to those found in the databases under the accession number AF159604 for the *P. westermani* strain Thailand. However, the sequences of P. westermani-like did not exhibit a striking similarity to any of those found in the databases.

DISCUSSION

In this study, we observed adult P. westermanilike samples obtained from a cat that was inoculated with the metacercariae. The adult had an ovary that was simply divided into six lobes, a seminal receptacle filled with spermatozoa, and cuticular spines arranged singly. These morphological features at the adult stage are in good agreement with the description of P. westermani (Thai strain) (Sugiyama et al, 2001; Binchai et al, 2005). With regard to the morphology of metacercariae, other than the size, the features of *P. westermani*-like were almost identical to those of P. westermani. Therefore, it can be concluded that *P. westermani*-like should be classified as P. westermani, or as one of the members (a cryptic species) of the P. westermani complex (Blair et al, 1997), based on the anatomical similarities.

We investigated the susceptibility of feline hosts to P. westermani-like by experimental infection, and compared the results with those of P. westermani. From the cat experimentally infected with P. westermani, worms were detected only in the lungs or pleural cavity. The worms recovered were identified as adults or at least pre-adults (Binchai et al, 2005). In contrast, as shown in this study, juvenile P. westermani-like lodged predominantly in the liver, while some matured into adults in the pleural cavity or lungs. These findings suggested that the susceptibility in cats differed between P. westermani and P. westermani-like. The susceptibility of feline hosts to P. westermani was also examined using worms from Malaysia (Habe et al. 1996). About half of the worms recovered were identified as juvenile worms, but the principal domicile of the juveniles was not the liver but the skeletal muscles.

Molecular comparison based on ITS2 sequences revealed that there were a few nucleotide differences (2.8%) between *P. westermani* (*P. westermani* strain Thailand) and *P. westermani*-like. Therefore, in order to determine the proper taxonomic status of *P. westermani*-like, we need to investigate the detailed phylogenetic relationships between *P. westermani*-like and *P. westermani*. In terms of the susceptibility of *P.* *westermani*-like, information regarding hostparasite relationships, particularly relating to the first intermediate hosts, is required. Studies into these issues are currently underway (Binchai *et al*, 2007).

REFERENCES

- Binchai S, Rangsiruji A, Ketudat P, Morishima Y, Sugiyama H. Morphological and genetic characterization of Thai *Paragonimus westermani* matured in cat. [Abstract]. The Joint International Tropical Medicine Meeting, 2005. Bangkok: Mahidol University, 2005:193.
- Binchai S, Rangsiruji A, Ketudat P, Morishima Y, Sugiyama H. Molecular systematics of a new form of *Paragonimus westermani* discovered in Thailand. *Southeast Asian J Trop Med Public Health* 2007;38 (suppl 1):92-6.
- Blair D, Agatsuma T, Watanobe T, Okamoto M, Ito A. Geographical genetic structure within the human lung fluke, *Paragonimus westermani*, detected from DNA sequences. *Parasitology* 1997;115: 411-7.
- Blair D, Waikagul J, Honzako Y, Agatsuma T. Phylogenetic relationships among the Thai species of *Paragonimus* inferred from DNA sequences. In: Tada I, Kojima S, Tsuji M, eds. Proceedings of the Ninth International Congress of Parasitology. Bologna: Monduzzi Editore, 1998:643-7.
- Bowles J, Blair D, McManus DP. A molecular phylogeny of the human schistosomes. *Mol Phylogenet Evol* 1995;4:103-9.
- Habe S, Lai KPF, Agatsuma T, Ow-Yang

CK, Kawashima K. Growth of Malaysian *Paragonimus westermani* in mammals and the mode of transmission of the fluke among mammals. *Trop Med Health* 1996;24:225-32.

- Miyazaki I. Paragonimiasis. In: Miyazaki I, ed. An illustrated book of helminthic zoonoses. Tokyo: International Medical Foundation of Japan, 1991:76-146.
- Naiyanetr P. Freshwater crabs in Thailand. Bangkok: Phaisalsipa Press, Bangkok. 1988:15 (Book published in memory of the royal cremation of Associate Professor Dr Praphun Chitchumnong of Chulalongkorn University).
- Rangsiruji A, Sugiyama H, Morishima Y, et al. A new record of Paragonimus other than P. westermani in southern Thailand. Southeast Asian J Trop Med Public Health 2006;37 (suppl 3):57-61.
- Sugiyama H, Sonoda J, Okuda M, Tomimura T. The macaque monkey as an experimental paratenic host for *Paragonimus westermani* (Kerbert, 1878) Braun, 1899. J Vet Med Sci 1984;46:345-56.
- Sugiyama H, Shibahara T, Ketudat P, Thaithong S, Kawashima K. Morphological re-examination of *Paragonimus westermani* described by Daengsvang and others in 1964. *Trop Med Health* 2001;29:371-4.
- Sugiyama H, Morishima Y, Kameoka Y, Kawanaka M. Polymerase chain reaction (PCR)-based molecular discrimination between *Paragonimus westermani* and *P. miyazakii* at the metacercarial stage. Mol Cell Probes 2002;16:231-6.