CASE REPORT

MEASLES-MUMPS-RUBELLA VACCINATION INDUCED THROMBOCYTOPENIA: A CASE REPORT AND REVIEW OF THE LITERATURE

Somchai Owatanapanich1, Nasamon Wanlapakorn2, Ratsamee Tangsiri1 and Yong Poovorawan2

1Department of Pediatrics, King Narai Hospital, Lop Buri; 2Center of Excellence in Clinical Virology, Faculty of Medicine, Chulalongkorn University, Bangkok, Thailand

Abstract. Immune thrombocytopenia (ITP) is a disease with autoimmune destruction of platelets. ITP among children has been associated with viral infections and some vaccinations. We report a case of ITP after measles-mumps-rubella (MMR) vaccination in a 10-month-old male infant who presented with purpura and acute gastrointestinal bleeding. This case was successfully treated with corticosteroids and intravenous immunoglobulin. ITP is a rare complication of the MMR vaccine that physicians should be aware of.

Keywords: measles-mumps-rubella, immunity, thrombocytopenia

INTRODUCTION

The definition of immune thrombocytopenia (ITP) according to the International Working Group is a platelet count <100 x 10^9/l with an unexplained cause (Journeycake, 2012). The annual incidence rate for ITP varies by country from 1.1 to 5.3 per 100,000 children (Terrell et al, 2010). The peak age-stratified prevalence of pediatric ITP is 2-5 years (Yong et al, 2010). Possible etiologies of secondary ITP include chronic infection (caused by Helicobacter pylori, human immunodeficiency virus and hepatitis C virus), acute infection (with Epstein-Barr virus, influenza virus and Varicella zoster virus), autoimmune disorders (such as systemic lupus erythematosus, antiphospholipid syndrome and rheumatoid arthritis), certain medications, and vaccinations (Johnsen, 2012). The majority of ITP cases associated with vaccinations are attributable to the measles-mumps-rubella (MMR) vaccine, the varicella vaccine, hepatitis A vaccine, and the tetanus-diphtheria-acellular pertussis (Tdap) vaccine (O’Leary et al, 2012). France et al (2008) identified 595 children aged <18 years with ITP and found a strong correlation between MMR vaccination and the risk for developing ITP in children aged 12-23 months. The absolute risk for developing ITP after the MMR vaccine varies by country, but is estimated to be between 0.087 and 4 (median 2.6) per 100,000 doses (Mantadakis et al, 2010).
The risk of severe ITP resulting from vaccination is low. No cases of ITP associated with MMR vaccine have yet been reported for Thailand. We report here the case of a 10-month old infant who developed thrombocytopenia with bleeding after receiving the MMR vaccine. The case was successfully managed with a therapeutic platelet transfusion, corticosteroids and intravenous immunoglobulin.

CASE REPORT

A well-nourished 10-month old Thai boy living in Lop Buri Province presented with pallor, dry lips, petechiae, and purpura extensively distributed over his body and oral mucosa. He had reportedly passed dark-colored stools for 3 days. The patient had received an MMR vaccine at King Narai Hospital 8 days prior to admission. He was afebrile with a heart rate of 120 beats/min and a blood pressure of 100/60 mmHg. Cardiovascular, respiratory and neurological examinations were unremarkable. Lymph nodes, liver and spleen were not palpable. A complete blood count revealed a hemoglobin level of 10.2 g/dl, a hematocrit of 30.3%, a white cell count of 6.64x10^9/l (31% neutrophils, 65% lymphocytes, 3% atypical lymphocytes, 1% monocytes, and no blast cells) and a platelet count of 8.5x10^9/l. A peripheral blood smear showed normochromic normocytic red cells, some polychromasia, no schistocytes, a normal white blood cells with predominant lymphocytes, and markedly decreased platelets. A coagulogram revealed a normal prothrombin time and a normal activated partial thromboplastin time. Renal and liver function tests were normal. Viral serologies (dengue NS-1 antigen, IgM, IgG, and anti-HIV) were all negative. No other cause of thrombocytopenia was detected. Therefore, the recent MMR vaccine was suspected as the cause of his thrombocytopenia. Because the patient had gastrointestinal bleeding from severe thrombocytopenia, intravenous immunoglobulin (IVIG) (1 g/kg/day) was administered daily for 2 days and intravenous dexamethasone at 6 mg/day was also given. On the second day of hospitalization, the patient’s hematocrit decreased to 24.9%, and the platelet count decreased to 8.0x10^9/l. He was transfused with 100 ml leukocyte-poor packed red blood cells and 1 unit of platelet concentrate. The following day, his platelet count had increased to 51.7x10^9/l and his hematocrit was stable. Three days after IVIG administration, the patient was no longer pale, and the petechiae and purpura had disappeared. He finally began passing yellow-colored stools which had previously been dark. His platelet count then increased to 100.9x10^9/l. The corticosteroid was discontinued on the fourth day of hospitalization. He was discharged home after a 5 day hospital stay with a platelet count of 165.8x10^9/l. He appeared healthy and had a normal platelet count at the follow-up visit (Fig 1).

DISCUSSION

This patient presented with an acute episode of gastrointestinal bleeding and severe thrombocytopenia eight days after MMR vaccination. After exclusion of other possible causes of thrombocytopenia, the diagnosis of MMR vaccine associated ITP was made. ITP is rarely caused by MMR and it is the only vaccine with a proven association with ITP (Cecinati et al, 2013). The pathogenesis is hypothesized to be the result of molecular mimicry and/or immune provocation by specific antigen exposure that activates...
the immune response to platelets in susceptible individuals (Johnsen, 2012). A systematic review of the literature found 12 studies from 10 countries showing the incidence of ITP after MMR vaccination ranges from 0.087 to 4 cases per 100,000 vaccine doses (Mantadakis et al., 2010). This variation may be due to different surveillance methods rather than a difference in the incidence, since the incidence was higher in countries with active surveillance systems compared to those with passive surveillance systems. MMR vaccine-related ITP usually occurs within 6 weeks of vaccination (Cecinati et al., 2013). In most cases, clinical symptoms are mild and may include bruising and petechiae, which resolve spontaneously within a few days to a few weeks (Cecinati et al., 2013). One study found that of all children diagnosed with acute ITP after MMR immunization, 93% of cases resolved within 6 months (Mantadakis et al., 2010). The development of chronic ITP after MMR vaccination and severe bleeding episodes in MMR-induced ITP are also rare (Kuhne et al., 2003). The case described here had severe thrombocytopenia with active gastrointestinal bleeding, which was treated with transfusion of packed red blood cells and platelets to prevent further complications.

The current guidelines of the Working Party of the British Committee for Standards in Haematology Task Force for initial management of children with acute ITP state that intervention is reserved for children with hemorrhage and a platelet count <20 x 10^9/l or for those with a life-threatening bleeding irrespective of the platelet count (Blanchette and Bolton-Maggs, 2008). The traditional regimen is
oral prednisolone at a dose of 2 mg/kg/day for 3-4 weeks. However, higher doses of steroids have been used and may result in a faster increase in platelet counts. Intravenous immunoglobulin, which blocks Fc-receptors on the macrophages of the reticuloendothelial system, is also the first line treatment for ITP in an emergency setting and proves effective when used at a high dose of 1-2 g/kg (Labarque and Van Geet, 2014). In this case, both intravenous immunoglobulin and high dose steroids were given to this patient due to severe bleeding and thrombocytopenia. As a result, his platelet level reached the treatment goal shortly thereafter. However, transient rebound thrombocytosis to $602 \times 10^9/\ell$ was observed eight days after IVIG administration. This thrombocytosis may have been due to the decrease in destruction of the opsonized platelets via the FcγR system, while the number of megakaryocytes in the bone marrow is usually normal or slightly elevated in ITP patients (Cooper et al, 2004).

In Thailand, two doses of the measles vaccine are required because only 76% of children are protected after the first dose at 9 months of age (Techasena et al, 2011). Moreover, unlike many other countries, the first dose of measles vaccine in Thailand is usually given at 9 months when maternal antibodies can interfere with the vaccine efficacy in some children (Albrecht et al, 1977; Schlereth et al, 2000). Here, we report a case of MMR vaccine associated ITP presenting with life-threatening bleeding. A question arose as to whether this child should receive a second dose of MMR vaccine. According to previous studies, the risk of recurrence of ITP after repeated doses of MMR vaccine is very low. Vlacha et al (1996) reported a case of recurrent ITP after the third dose of MMR vaccine. A systematic review showed among 26 children with a history of MMR vaccine induced ITP, none developed recurrence of ITP after the second dose of MMR (Mantadakis et al, 2010). The Immunization Monitoring Program ACTive (IMPACT) in Canada recommends measuring measles antibody titers in children with MMR vaccine related ITP before booster administration in order to decide whether a second dose is required (Sauvé and Scheifele, 2009). Since measles remains endemic in Thailand, we recommend a second dose of MMR vaccine in children with a history of MMR vaccine induced ITP due to the low risk of recurrence of ITP and the higher risk of natural measles infection; the benefits of vaccination outweigh the risk of severe thrombocytopenia associated with immunization. Nevertheless, patient education is important to raise awareness of the side effects of vaccination. Surveillance of platelet counts may be warranted in children with a history of bleeding following MMR vaccine administration.

ACKNOWLEDGEMENTS

The authors wish to thank Dr Veerasak Kronglarpcharoen and Dr Weerapat Owatanapanich for their helpful comments and Dr Sompong Yongpunsawad for reviewing the manuscript. We also thank the National Research University Project, Office of Higher Education Commission (HR1155A, WCU-001, 007-HR57), the Center of Excellence in Clinical Virology, Chulalongkorn University, the Ratchadaphiseksomphot Endowment Fund of Chulalongkorn University (RES560530093) the Outstanding Professor of the Thailand Research Fund (DPG5480002); and King Chulalongkorn Memorial Hospital Thai Red Cross Society, MK Restaurant Company Limited and The Siam Cement Pcl for supporting Prof Yong Poovorawan.
REFERENCES


