IMMUNOLOGICAL STUDY OF TYPHOID FEVER IN MAN: HUMORAL IMMUNE RESPONSE

VARANYA SANGPETCHSONG and SAVANAT THARAVANIJ

Department of Microbiology, Faculty of Public Health and Department of Microbiology and Immunology, Faculty of Tropical Medicine, Mahidol University, Bangkok 4, Thailand.

INTRODUCTION

It has been generally accepted that cell mediated immunity plays an important role in the recovery from typhoid fever (Ushiba, 1965; Blanden et al., 1966). Nevertheless the possible protective role of humoral antibodies have not been excluded (Ornellas et al., 1970). Humoral antibodies are usually detected by the Widal test, the sensitivity of which is not yet satisfactory, since false negative was found in 24-34 % of patients (Stuart and Pullen, 1946; Chompoosang et al., 1977). The other test is the indirect haemagglutination using either alkaline treated lipopolysaccharide or Vi antigen (Schubert et al., 1959; Chernokhvostova et al., 1969). The course of humoral immune response in patients with typhoid fever has been reported (Kumar et al., 1974; Balakrishna et al., 1977). The prominant immunoglobulin class of the antibody in patients with typhoid fever was predominantly IgM (Chernokhvostova et al., 1969; Kumar et al., 1974). Since indirect haemagglutination test has been shown to be more sensitive than the Widal test in experimental animal immunised with 0 somatic antigen (Carlsson et al., 1972), studies were carried out to determine whether this finding is also applicable to typhoid fever in man. The study was designed also to cover the time course of the haemagglutinating antibodies response and the determination of the immunoglobulin class responsible for this antibody activity.

MATERIALS AND METHODS

Bacterial strains: Salmonella typhi 0901 and H901 obtained from the Faculty of Tropical Medicine, Mahidol University and the Department of Medical Science, Ministry of Public Health respectively were used throughout the study.

Subjects: The study was conducted in 45 patients with typhoid fever, whose ages ranged from 15 to 42 years. Diagnosis of these patients was done either by demonstration of positive blood culture (35 patients) or the presence of high 0 agglutinating antibody titer ($\geq 1:320$) in a single serum specimen (10 patients). In addition 4 patients with paratyphoid A fever were studied. The controls included 26 healthy volunteers and 14 hospitalized patients 13 of whom had pyrexia of unknown origin (PUO) and the remaining one had scrub typhus.

Widal test: The classical tube method of Widal 0 and H agglutination was used (Bailey and Scott, 1974). The optical density of the bacterial suspension was adjusted to give 0.21 at 650 nm as measured by Coleman Junior II spectrophotometer which was equivalent to 9×10^8 CFU per ml. The antigen was found to give the same titer as the Widal 0 antigen from Difco (Batch no. 2842-56, control 623285) when tested against the same sera.

Indirect haemagglutination (IHA) test:The LPS used to sensitize sheep red blood cell (SRBC) was prepared from *S. typhi* 0901 according to the technique of Neoh and Rowley (1970). The LPS was rendered alkaline by treatment with NaOH according to the technique of Auzins (1968). The microtiter technique of indirect haemagglutination modified from the macrotechnique originally described by Auzins (1968) was used. The immunoglobulin class of the IHA antibody was determined by the anti-immunoglobulin enhancement as described by Steel *et al.*, (1974) with minor alteration in that 2.5%SRBC suspension was used instead of 1% cell suspension. Preliminary testing showed that enhancement of IHA titer was optimum at the anti-immunoglobulin (Kallestad, Minnesota) dilution of 1:2000. This dilution was then chosen for use throughout the four week course of this study.

RESULTS

Result of serum antibody titers in 45 patients with typhoid fever and in the controls are presented in Table 1. The patients with typhoid fever had significant higher Widal 0 agglutinin, H agglutinin titers than those of the health individuals and febrile control. Twenty-three paired sera taken 7-14 days apart were available for testing. There was no statistically significant difference between the titers of the first and second samples.

Time course humoral responses from 72 specimens in 45 patients is shown in Fig. 1. In comparison with healthy control, there was a significant increase in the Widal 0 and H titers and IHA titers in the second week of illness and thereafter (p < 0.001). In addition,

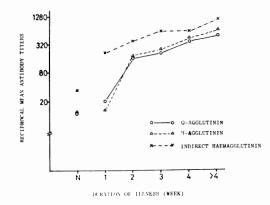


Fig. 1—Means of reciprocal 0 agglutinins, H agglutinins and indirect hemagglutinins in normal (N) and in patients with typhoid fever at various time of illness.

the IHA antibody titers were slightly higher than Widal 0 and H antibody titers in all stages of the tisease.

Analysis of variance of the antibody response at different time of illness revealed that there was a significant rise of Widal 0 and H antibody titers (p < 0.05), but not the IHA titer (p>0.05). Further analysis by Duncan's new multiple range test showd that significant rise in titer was found only between sera the of first week and those of subsequent weeks.

Correlation between Widal 0 agglutinin and IHA titers in 72 specimens from 45 patients

Subjects	No.	Geometrie	c mean O titer	Mean H titer	Student t test	
Subjects	tested	O agglutinin	Haegmagglutinin	Mean n mei	Student t test	
Typhoid	45	340.34	565.80	320	1 vs 2 p < 0.001	
Healthy control	26	10.72	31.62	11.48		
Fever control	14	10.47	25.12	11.75	1 vs 3 p < 0.001	

 Table 1

 Reciprocal peak natibody titers in typhoid patient and control.

Student 't' test 1 vs 2 = p < 0.001; 1 vs 3 = p < 0.001.

with typhoid fever showed very significant correlation (Fig. 2., r = 0.66; $p < 10^{-7}$)

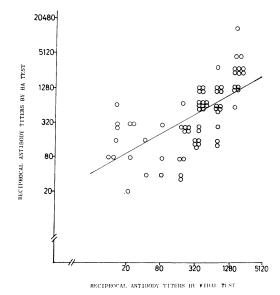


Fig. 2—Relationship between serum antibody titers measured by conventional Widal test and indirect hemagglutination test.

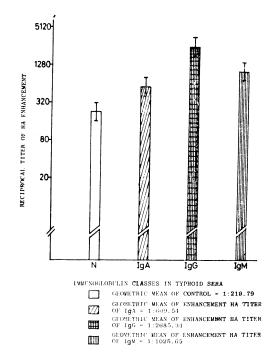
When comparison was made between the IHA and Widal agglutinating titers, it was found that there was significant difference as assessed by Wilcoxon matched-pairs signed-ranks test (p < 0.01) indicating that the antibody titer measured by IHA test was more sensitive than the Widal test.

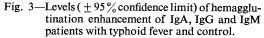
The immunoglobulin class of IHA antibody was determined by the immunoglobulin enhancement test. Enhancement of four fold rise or more of haemagglutinating titers was considered significant. The immunoglobulin class in 45 patients with typhoid fever was found to belong to 3 main classes of IgG, IgM and IgA (Table 2). Strongest enhancement of IgG was found in most patients (Fig. 3). Distribution of immunoglobulins in relation to duration of illness is shown in Table 3. It was observed that during the first week of illness only IgG and IgM but not IgA were detectable. In subsequent weeks all three immunoglobulin classes were elevated.

Table 2

The percentage of immunoglobulin antibodies in sera of typhoid patients.

Immunoglobulin class	Percentage
IgG, IgM and IgA	38.89
IgG and IgM	48.61
IgG and IgA	4.16
IgM	2.77
IgG	5.56





Natural IHA antibody against 0 antigen was demonstrated in all 22 healthy individuals but the titer was low (1:10-1:40). Immunoglobulin enhancement test in these sera showed that the antibody activity was associated chiefly with IgG and IgM.

HUMORAL IMMUNE RESPONSE IN TYPHOID FEVER

Table 3

Duration	Relative distribution of immunoglobulin antibody							
Duration — of illness	IgA, IgG and IgM	IgG and IgM	IgG and IgA	IgG	IgM			
1 st week(2)*	-	100.0	-	-	-			
2 nd week(26)	34.6	50.0	7.7	3.9	3.9			
3 rd week(16)	18.7	56.3	-	18.8	6.3			
4 th week(24)	50.0	45.8	4.2	-	-			
more than(4) 4 week	50.0	50.0	-	-	-			

Relative distribution of immunoglobulin in sera of typhoid patients.

*Number of cases investigated.

DISCUSSION

The time course development of antibodies in patients with typhoid fever showed that the antibodies appeared on the first week of illness with the gradual increase in titer thereafter. This finding was similar to those reported by other investigators (Kumar et al., 1974; Balakrishna et al., 1977). Comparison between IHA and Widal test by Wilcoxon matched-pairs signed-ranks test showed that the IHA test was more sensitive than the Widal test. This contention was further substantiated by demonstration of lower antibody IHA titers in healthy unvaccinated persons. False negative Widal test in patients with typhoid fever has been reported in 34.17% of 360 cases (Stuart and Pullen, 1946), and in 24.2% of Thai patients with typhoid fever (Chompoosang et al., 1977). In our studies, false negative Widal reaction was found only in 4 patients (8.9%) whose blood was taken during 12-19 days after the onset of clinical illness. The lower percentage of false negativity in our patients is not known. Since no detailed information was available with respect to the time course of clinical illness when sera were taken in the reports by Stuart and Pullen (1946) and by Chompoosang et al., (1977) it was likely that some of the negative sera in these patients were taken in the early state of illness. The prior treatment with antibiotics was unlikely to affect significantly the subsequent Widal antibody response since raised Widal antibody titer was found in most patients. Such contention would be in corroboration with those of Robertson et al., (1970) and Balakrishna et al., (1977). False negative IHA was found in 3 patients indicating that specificity of Widal and IHA tests were similar.

Antibodies to somatic antigen of S. typhi were found in all three major immunoglobulin classes. The results were concordant with the report of Chernokhvostova et al., (1969). However, our results showed that both IgG and IgM antibodies were produced in the majority of cases. This data contradicted that of Pinto and Dammaco (1964), who reported that the entirely IgM response were observed in patients with typhoid fever, as well as in healthy individuals receiving Salmonella vaccine (Chernokhvostova et al., 1969). It was also shown by Kumar et al., (1974), that antibody in typhoid fever was 2 mercaptoethanol (2 ME) sensitive and hence would presumably be IgM. However, they

reported that during the third week or more of illness, significant levels of anti-0 and anti-H antibodies resistant to 2 ME (IgG) developed which would indicate the development of other classes of antibodies besides IgM.

The strongest enhancement of haemagglutinating activity in the I2G class in most patients seems to be a paradox when taking into account that bacterial lipopolysaccharide antigen stimulates as a rule a prolonged and intensive synthesis of IgM antibody (Franklin 1964). It is possible that LPS antibody produced in patients is incomplete and its activity will be enhanced when anti-immunoglobulin is added. This type of incomplete (7S) antibody was demonstrated in typhoid carriers (Chernokhvostova et al., 1969). In addition the natural agglutinating antibodies to the 0 antigen of S. typhimurium was shown by Robbins et al., (1965) to belong to the IgG class when antiglobulin serum was added.

SUMMARY

Widal and indirect haemagglutination test were used to study the humoral immune response of 45 patients with typhoid fever. The Widal and IHA tests were positive in 41 (91.1%) and 42 (93.3%) of patients with typhoid fever after the second week of Comparison between the Widal illness. and IHA test showed that the latter was more sensitive as assessed by Wilcoxon matchedpairs signed-ranks test (p < 0.01). The anti-LPS antibody belongs to all three classes of immunoglobulins namely IgG, IgM and IgA with the confinement mostly to IgG and IgM. It was found that the level of haemaggluti nating enhancement activity was strongest in IgG class. The low level of anti LPS-antibody was found in healthy persons with the pattern of immunoglobulin class of antibody similar to that in patients with typhoid fever.

ACKNOWLEDGEMENTS

The authors would like to thank Dr. Danai Bunnag, Department of Clinical Tropical Medicine, Faculty of Tropical Medicine; Dr. Kraiwan Ruengvisuthi Department, of Medicine, Rajvithi Hospital and Dr. Wichain Soonthornsiri, Unit of Infectious Disease and Tropical Medicine, Police Hospital in providing specimens. Special thanks to Dr. Wanpen Chaicumpa for her advice.

REFERENCES

- AUZINS, I., (1968). A comparative assay of O-somatic antigen 5 of salmonellae. Aust.
 J. Exp. Biol. Med. Sci., 46:93.
- BAILEY, W.R. and SCOTT, E.G., (1974). Diagnostic Microbiology. 4th ed. The C.V. Mosby Company. Saint Louis, U.S.A., pp. 342.
- BALAKRISHNA SARMA, V.N., MALAVIYA, A.N., KUMAR, R., GHAI, P.O. and BAKHTARY, M.M., (1977). Development of immune response during typhoid fever in man. *Clin. Exp. Immun.*, 28 : 35.
- BLANDEN, R.V., MACKANESS, G.B. and COLLINS, F.M., (1966). Mechanisms of acquired resistance in mouse typhoid. J. Exp. Med., 124 : 585.
- CARLSSON, H.E., LINDBERG, A.A. and HAMMARSTROM, S., (1972). Titration of antibodies to Salmonella 0 antigens by enzyme-linked immunosorbent assay. *Infect. Immun.*, 6 : 703.
- CHERNOKHVOSTOVA, E., LUXEMBERG, K.I., STARSHINOVA, V., ANDREEVA, N. and GERMAN, G., (1969). Study on the production of IgG, IgA and IgM antibodies to somatic antigens of *Salmonella typhi* in human. *Clin. Exp. Immun.*, 4 : 407.
- CHOMPOOSANG, C., RUNGPITIRANGSI, B. and KONGSAMRAN, S., (1977). The relation between Widal agglutinin titers and haemoculture. *Siriraj Hosp. Gaz.*, 29:1779.

- FRANKLIN, E.C., (1964). The immune globulin their structure and function and some technique for their isolation. *Progr. Allergy.*, 8 : 58.
- KUMAR, K., MALAVIYAA, N., MURTHY, R.G.S., VENKATARAMAN, M. and MOHAPATRA, L.N., (1974). Immunological study of typhoid : Immunoglobulins, C_3 , antibodies and leukocyte migration inhibition in patients with typhoid fever and TAB-vaccinated individuals. *Infect. Immun.*, 10 : 1219.
- NEOH, S.H. and ROWLEY, D., (1970). The antigens of *Vibrio cholerae* involved in the vibriocidal action of antibody and complement. J. Infect. Dis., 121: 505.
- ORNELLAS, E.P., ROANTREE, R.J. and STEWARD, J.P., (1970). The specificity and importance of humoral antibody in the protection of mice against intraperitoneal challenge with complement sensitive and complement resistant salmonella. J. Infect. Dis., 121: 113.
- PINTO, L. and DAMMACO, F., (1964). Quoted by Chernokhvostova, E., Luxemberg, K.I., Starshinova, V., Andreeva, N. and German, G., (1969). Study on the production of IgG, IgA and IgM antibodies to somatic

antigens of Salmonella typhi in human. Clin. Exp. Immun., 4 : 407.

- ROBBINS, J.B., KENNY, K. and SUTER, E., (1965). The isolation and biological activities of rabbit γ M and γ G anti-Salmonella typhimurium antibodies. J. Exp. Med., 122: 385.
- ROBERTSON, R., FATHY, M. and WAHAB, A., (1970). Influence of chloramphenicol and ampicillin on antibody response in typhoid and paratyphoid fever. *Ann. Intern. Med.*, 72: 219.
- SCHUBERT, H.J., EDWARDS, R.P., and RAMSEY, H.C., (1959). Detection of typhoid carriers by agglutination tests. *J. Bact.*, 77 : 648.
- STEELE, E.J., CHAICUMPA,W. and ROWLEY, D., (1974). Isolation and biological properites of three classes of rabbit antibody to *Vibrio cholerae. J. Infect.*, *Dis.*, *130*:93.
- STUART, B.M. and PULLEN, R.L., (1946). Typhoid clinical analysis of 360 cases. *Arch. Intern. Med.*, 78 · 629.
- USHIBA, D., (1965). The types of immunity in experimental typhoid: "cellular immunity" and "humoral immunity". *Keio. J. Med.*, *14*: 45.